
































MEETING REPORT OPEN ACCESS

Dealing With the Complexity of Effective Population Size in Conservation Practice

Ancuta Fedorca^{1,2}  | Joachim Mergeay^{3,4}  | Adejoke O. Akinyele⁵  | Tamer Albayrak^{6,7}  | Iris Biebach⁸  | Alice Brambilla^{8,9}  | Pamela A. Burger¹⁰  | Elena Buzan^{11,12}  | Ino Curik^{13,14}  | Roberta Gargiulo¹⁵  | José A. Godoy¹⁶  | Santiago C. González-Martínez¹⁷  | Christine Gossen¹⁸  | Myriam Heuertz¹⁷  | Sean Hoban^{19,20}  | Jo Howard-McCombe²¹  | Maria Kachamakova²²  | Peter Klinga^{23,24}  | Viktoria Köppä²⁵  | Elenora Neugebauer^{26,27}  | Ivan Paz-Vinas²⁸  | Peter B. Pearman^{29,30,31}  | Laia Pérez-Sorribes¹⁶  | Baruch Rinkevich³²  | Isa-Rita M. Russo³³  | Adélaïde Theraroz¹⁷  | Nia E. Thomas³³  | Marjana Westergren³⁴  | Sven Winter^{10,35}  | Linda Laikre²⁵  | Alexander Kopatz³⁶ 

Correspondence: Alexander Kopatz (alexander.kopatz@nina.no)

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ABSTRACT

Effective population size (N_e) is one of the most important parameters in evolutionary biology, as it is linked to the long-term survival capability of species. Therefore, N_e greatly interests conservation geneticists, but it is also very relevant to policymakers, managers, and conservation practitioners. Molecular methods to estimate N_e rely on various assumptions, including no immigration, panmixia, random sampling, absence of spatial genetic structure, and/or mutation-drift equilibrium. Species are, however, often characterized by fragmented populations under changing environmental conditions and anthropogenic pressure. Therefore, the estimation methods' assumptions are seldom addressed and rarely met, possibly leading to biased and inaccurate N_e estimates. To address the challenges associated with estimating N_e for conservation purposes, the COST Action 18134, Genomic Biodiversity Knowledge for Resilient Ecosystems (G-BiKE), organized an international workshop that met in August 2022 in Braşov, Romania. The overarching goal was to operationalize the current knowledge of N_e estimation methods for conservation practitioners and decision-makers. We set out to identify datasets to evaluate the sensitivity of N_e estimation methods to violations of underlying assumptions and to develop data analysis strategies that addressed pressing issues in biodiversity monitoring and conservation. Referring to a comprehensive body of scientific work on N_e , this meeting report is not intended to be exhaustive but rather to present approaches, workshop findings, and a collection of papers that serve as fruits of those efforts. We aimed to provide insights and opportunities to help bridge the gap between scientific research and conservation practice.

For affiliations refer to page 13.

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1 | Applicable Methods for Estimating Effective Population Size Are Needed

Effective population size (N_e), defined as the size of an ideal population that experiences the same amount of genetic drift and increase of inbreeding as the real population (Wright 1931), is one of the most important parameters for assessing the long-term viability of species and is, therefore, an important measure of conservation biology. The effective size of a population is essentially an evolutionary analogue to the census size (N_c), and it is a quantity that correlates to the loss or maintenance of genetic diversity and inbreeding within a population (Waples 2024a; Waples 2022). Higher N_e results in more maintenance of genetic diversity or lower levels of inbreeding and a faster response to natural selection and, thereby, adaptation to environmental changes. Hence, populations with higher N_e are expected to have higher survival probability. In 2022, due to the general acknowledgement of its importance for biodiversity conservation, N_e became the basis for a headline indicator for the monitoring and reporting of genetic diversity under the monitoring framework of the UN's Convention on Biological Diversity (CBD) Kunming-Montreal Global Biodiversity Framework (GBF) (i.e., headline indicator A.4; CBD 2022; Hoban, da Silva, et al. 2024; Hoban, da Silva, et al. 2023). Consequently, effective population size is now embraced by governmental bodies and policy stakeholders, including national focal points for the CBD. In addition, effective population size has been included in relevant Essential Biodiversity Variables (EBV) for Genetic Composition (Hoban et al. 2022). Thus, practical and easy-to-use tools are needed to allow a diverse group of users to monitor and report progress on effective population size (Mastretta-Yanes et al. 2024). However, choosing which method to apply and how to interpret the results is not straightforward. Given the different types of effective population sizes (Box 1), all referred to as N_e , the plethora of methods to calculate them, and the increasing number of different data sources available, we believe that there is a lack of scientifically evaluated and harmonized guidance for global, national, and regional reporting of N_e .

Depending on how a particular study is designed, from sampling to the analysis method used, estimates of N_e from the same population can vary by orders of magnitude, not least

BOX 1 | What is N_e ?

Effective population size (N_e) is defined as the size of an ideal population that experiences the same amount of a given genetic property as the real population. In its purest sense, it assumes that a population is isolated and is at mutation-drift equilibrium. There are many different types of N_e (inbreeding, variance, additive variance, eigenvalue, coalescence, metapopulation N_e), which are identical when the population is closed and at mutation-drift equilibrium. We refer to Ryman, Laikre, and Hössjer (2019) for a comprehensive overview of how these differ when these conditions are not met. In the context of conservation genetics, the genetic properties ideally used to define N_e are additive variance N_e , allele frequency variance N_e , or inbreeding N_e , but other properties such as coalescence or linkage disequilibrium N_e can also be used. See our glossary (Box 2) for the definitions of these properties.

because different types of N_e (Box 1) have different meanings in space (Ryman, Laikre, and Hössjer 2019; Waples 2024a) and time (Nadachowska-Brzyska, Konczal, and Babik 2022; Tenesa et al. 2007) (see Figure 1). Different population properties affect N_e estimates, even when these converge to the same value under random sampling and certain ideal theoretical conditions, i.e., the population is isolated, of constant size, panmictic, has non-overlapping generations, and is in mutation-drift equilibrium (Ryman, Laikre, and Hössjer 2019; Waples 2024a). However, the lack of differentiation among different types of effective population size, often indistinguishably termed N_e , leads to confusion (e.g., Fady and Bozzano 2021; Hoban, Paz-Vinas, et al. 2021).

The definition and, consequently, the value of N_e will also depend on the spatial extent of the target population and the sampling scheme. Populations can be isolated or connected by gene flow, forming metapopulations. When populations are not isolated, the administrative boundaries often do not coincide with the biological population, and it can be difficult to determine their size. Sampling schemes can involve several interconnected populations, a single population regularly sampled (connected or not with others), or a portion of a larger continuous population (Box 2).

On the temporal scale, N_e can refer to historical N_e or contemporary N_e . Historical N_e is a geometric mean of N_e per generation over many generations. As such, it explains the current genetic make-up of a population and can be difficult to link with historical events, such as past management or anthropogenic environmental change. Contemporary N_e indicates the N_e of the current generation (or a few previous generations) and reflects the drift to be expected in the near future. This is of relevance for the ongoing monitoring of populations.

In real-world conservation management, where formerly large populations have often become fragmented into small subpopulations, almost none of the underlying assumptions (i.e., isolation, panmixia, constant population size, etc.) are met (Ryman, Laikre, and Hössjer 2019). Thus, estimates of N_e obtained under realistic circumstances may be more or less inaccurate. Moreover, for practical conservation, N_e estimates may not be what nature managers, policymakers, and researchers believe they represent at the spatial and temporal scales. At the spatial scale, depending on how sampling was conducted relative to the actual population range and the method used, one could be estimating the N_e of a part of the population (subpopulation) or of the entire metapopulation (see Figure 1A and Ryman, Laikre, and Hössjer 2023; Waples 2010). When estimating N_e for conservation, it is essential to first determine the spatial scale of the ancestral and/or current metapopulation, as overlooking the importance of the spatial scale leads to dubious results (Clarke et al. 2024). As management units, sometimes determined by political boundaries, rarely correspond completely to biological populations, N_e estimates may not accurately reflect the status of the assessed populations and species, potentially misleading conservation planning, decisions, and actions. At the temporal scale, and depending on the estimation method, N_e estimates might reflect the historical N_e across several, up to hundreds of generations (i.e., coalescent N_e), or, when using methods for calculating contemporary N_e , the N_e estimate obtained might represent the N_e in the last two to three generations (Nadachowska-Brzyska, Konczal, and Babik 2022).

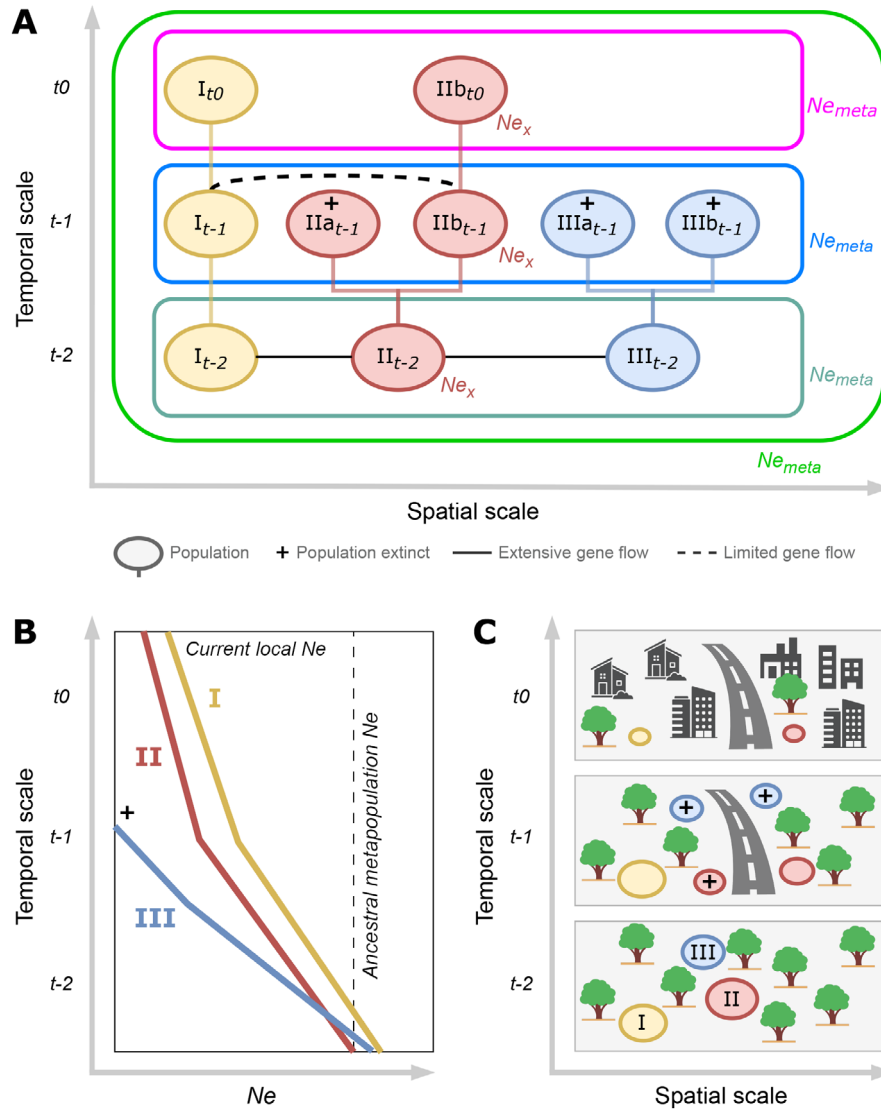


FIGURE 1 | Schematic representation of a metapopulation evolving through time as an example, intended to highlight the possible ambiguity of Ne estimations. The X-axis indicates populations' spatial distribution (sites) along one spatial dimension, for simplicity. As we go up and forward in time (Y-axis), subpopulations disappear, and others are formed, but the metapopulation as a whole is maintained. (A) Sampling a single subpopulation (IIb) in the present (t_0) and applying different Ne estimation methods may result in vastly different Ne estimates representing different aspects of the effective size, which are all commonly called “ Ne ”. Disambiguation of these different meanings is essential in conservation. Depending on the approach, one can estimate from the same sample local contemporary N_{ex} (e.g., using linkage disequilibrium, kinship, or a temporal method), contemporary Ne_{meta} (when based on heterozygosity (He) decay across time; this requires two samples across time; pink), coalescent Ne (when based on its current He , and assuming mutation-drift equilibrium; chartreuse), Ne_{meta} at different times in the past (blue, seagreen), but never past N_{ex} . (B) Methods that estimate recent Ne trajectories (0–200 generations ago) will initially reflect local Ne , but will increasingly reflect metapopulation Ne , and samples taken in different subpopulations but with origins in the same metapopulation will eventually converge on the same Ne_{meta} , which is the sum of the past N_{ex} (here at $t-2$). The risk is that this is interpreted as a population decline, whereas it represents a confounding effect of spatial scale (Novo et al. 2023). (C) A landscape-level schematic depiction of the processes occurring in (A) and (B).

Despite the pronounced scientific knowledge-to-application gap, genetic diversity concepts are increasingly being integrated into mainstream conservation, management, and biodiversity policy (Bertola et al. 2024; Hoban, Bruford, et al. 2021), where Ne remains a crucial summary statistic to evaluate the long-term survival capacity of natural populations (Hoban, Paz-Vinas, et al. 2021). Our article describes a workshop focused on refining methods for testing Ne using real-world datasets. Attendees collaborated to address challenges such as data availability, missing data, and testing barriers across various taxa. The key outcome was to align conservation theory with practical

challenges by estimating population sizes (N_c , Ne), focusing on species of conservation concern, and including diverse life histories and taxonomic groups to maximize conservation impacts.

1.1 | Complexity and Reality of Ne Estimates

Over the past decade, the accessibility of DNA-based and genetic monitoring has increased due to declining sequencing costs, wider availability of genomic data across many species, capacity-building endeavors, and investments from international (e.g.,

BOX 2 | Glossary.

Additive variance—Total effect on a trait stemming from one or more gene loci. **Census size (N_c)**—Number of individuals in a population that are reproductively mature. **Coalescence**—In population genetics, a model of how alleles sampled from a population may have originated from a common ancestor. **Demographic bottleneck**—an event that drastically reduces the census size of a population. **Effective population size (N_e)**—Size of an ideal population that experiences the same rate of genetic drift as the observed population. **Genetic drift**—Random sampling of allelic variants that may lead to changes in the frequency of existing alleles from one generation to the next due to chance. **Isolation by distance (IBD)**—The decrease in the genetic similarity among individuals or populations as the geographic distance between them increases. **Life-history traits**—A set of coevolved traits that affect an individual's survival and reproductive potential. **Linkage disequilibrium (LD)**—The nonrandom association of alleles at different loci. **Metapopulation**—In population genetics, a group of spatially separated populations of the same species that are connected by gene flow. **Mutation-drift equilibrium**—The balance between new mutations introducing genetic diversity and random genetic drift removing (fixing) variants in a population. **Panmixia**—Random mating of individuals within a population that results in equal parental contributions to the next generation. Under Hardy-Weinberg assumptions, random mating occurs when allele frequencies accurately predict genotype frequencies.

EU), national, and private initiatives in conservation genomics (e.g., Earth Biogenome Project, EBP; International Barcode of Life, iBOL; or European Reference Genome Atlas, ERGA) (Theissinger et al. 2023). It has become practical and affordable to genotype individuals for tens of thousands of markers and estimate N_e with confidence intervals. However, numerous theoretical, technical, and methodological issues when estimating N_e need to be considered (Cox, Neyrinck, and Mergeay 2024; Gargiulo, Decroocq, et al. 2024; Mergeay et al. 2024; Pérez-Sorribes et al. 2024; all in this Special Issue *Effective population size in conservation and biodiversity monitoring*) depending on the specific genetic markers, analysis methods, and sampling schemes.

Conservation genetic studies aimed at estimating allele frequencies have typically sampled 30–50 individuals, allowing the calculation of useful summary statistics (Allendorf 2017). When multiple sites are sampled, F -statistics might be used to infer the genetic structure among subpopulations. N_e can be estimated from such samples using single-sample estimator approaches (Jones and Wang 2010; Waples and Do 2010). These methods, however, can be sensitive to violations of the model assumptions, potentially leading to seemingly precise N_e estimates (i.e., with narrow confidence intervals) that may, however, be highly biased (Nunney 2016; Ryman, Laikre, and Hössjer 2023; Waples 2010).

Existing methods for N_e estimation vary widely in what they intend to estimate (N_e trajectories vs. point estimates; historical vs. contemporary N_e), the genetic signal they capture, and the

data they require. Table 1 provides a brief description of the most commonly used methods, along with their main caveats. Most methods will assume neutrality and an isolated, random mating population with discrete generations. Population structure, in particular, biases N_e estimates obtained by many methods (Chikhi et al. 2010).

For a more complete description of available methods and detailed accounts of their associated temporal aspects, see Nadachowska-Brzyska, Konczal, and Babik (2022).

Here, we use the terminology of Ryman, Laikre, and Hössjer (2019) to designate specific aspects of N_e . We distinguish between the N_e of a subpopulation x in isolation (N_{ex}) and the realized N_e of subpopulation x when the joint effects of drift and gene flow are taken into account (N_{eRx}), metapopulation N_e (N_{eMeta}), and coalescent N_e (N_{eCo}), based on frequently used methods for N_e estimation. Although important in their own way, we do not explore the differences among variance N_e , inbreeding N_e , gene diversity N_e , the effective number of breeders N_b , or additive variance N_e (Ryman, Laikre, and Hössjer 2019; Waples 2005). Notably, the common case of isolation by distance (IBD) in continuous populations can be considered as the ‘neighbourhood’ effective size ($=N_{ex}$ of the neighbourhood) if the samples originate from a single neighbourhood (Neel et al. 2013; Cox, Neyrinck, and Mergeay 2024). N_{eCo} can be seen as the long-term N_e that reflects the gene diversity under the assumption of mutation-drift equilibrium.

To illustrate the ambiguity of these different types of N_e , a simplified example is provided in Figure 1A, where we consider a population consisting of two isolated subpopulations (I and IIb), with each subpopulation experiencing random mating. Suppose that subpopulation IIb is sampled at time t_0 ; we could estimate N_{ex} from linkage disequilibrium (LD) (Waples and Do 2008) or sibship frequency (Jones and Wang 2010). With two samples from subpopulation IIb across a certain time span, we could also calculate (the variance) N_{ex} using a temporal method, or at least the harmonic mean across the sampled time span. Under some circumstances, we could estimate N_{eRx} using a temporal method that calculates the net genetic drift over that time span considering gene flow (Ryman, Laikre, and Hössjer 2019, but see Nunney 2016). Although at migration-drift equilibrium, N_{eRx} is similar across subpopulations and of the same magnitude as N_{eMeta} , N_e values obtained from the same subpopulation can easily be an order of magnitude different, yet they are all called “the effective size” (see Figure 1A–C).

It is possible to estimate the change in N_e over time for a specific sample using methods that trace back the historical trajectory of N_e over recent time (10s to 100s of generations) or ancient times (10^3 to 10^5 generations ago) (Nadachowska-Brzyska, Konczal, and Babik 2022). When making inferences over time, we also need to acknowledge the limitations of changes in the spatial scale of reconstructions. While we sample an individual at time t , we also sample half of its parents’ and a quarter of each of its grandparents’ genes. We are also sampling across a wider geographic area as the spatial origins of ancestors of each individual widen the spatial scale due to gene flow across ancestral subpopulations. Therefore, when we sample a subpopulation and get an estimate of its N_e across time, it becomes increasingly difficult

TABLE 1 | Some methods commonly used to estimate effective population size N_e .

Estimation	Time scale	Type of method	Methods	Data required	Caveats	References
Effective population size (N_e) trajectories	Historical	Coalescent-based	Skyline plots	DNA sequences	N_e reflects historical N_e of the total population. Sensitive to past population structure, which is often unknown.	Drummond et al. 2005; Ho and Shapiro 2011
	Historical	Sequentially Markovian Coalescent (SMC)	PSMC, MSMC	Single (PSMC) or multiple (MSMC) genomes sequenced to high coverage	Population structure can create spurious signals of population size change. Sampling regime can be important.	Li and Durbin 2011; Schiffels and Durbin 2014; Schiffels and Wang 2020
	Historical	Site frequency spectrum (SFS)	Stairway plot	Folded/unfolded SFS from hundreds of samples	May confound changes in gene flow and demographic changes.	Liu and Fu 2015, 2020
	Recent	Linkage Disequilibrium (LD)	GONE, LinkNe, SNeP	Phased or unphased genome-wide genotype data	Will initially estimate local N_e and increasingly estimate metapopulation N_e , unless the population has always been closed.	Barbato et al. 2015; Hollenbeck, Portnoy, and Gold 2016; Santiago et al. 2020, 2024

(Continues)

TABLE 1 | (Continued)

Estimation	Time scale	Type of method	Methods	Data required	Caveats	References
Effective population size (N_e)	Long-term	Site frequency spectrum (SFS)	dadi, fastsimcoal2	Folded/unfolded SFS from tens to hundreds of samples	May confound changes in gene flow and demographic changes.	Gutenkunst et al. 2009; Excoffier et al. 2013; Excoffier et al. 2021
	Contemporary	Linkage Disequilibrium (LD)	LDNe, NeEstimator V2	Genotype data from a single time-point sample	Assumes independent markers, discrete generations, closed populations, and random mating. Very sensitive to spatial genetic structure.	Do et al. 2014; R. S. Waples and Do 2008
	Contemporary	Heterozygosity Excess	NeEstimator V2	Genotype data from a single time-point sample	Assumes the only source of Hardy-Weinberg deviations is local inbreeding. Rarely accurate when N_e is not very small. To be avoided.	Balloux 2004; Do et al. 2014
	Contemporary	Temporal	NeEstimator V2; NB (R package), decay of heterozygosity over time	Genotype data from two or more samples separated by a known number of generations	Avoids confusing spatial and temporal genetic structure when sampling in metapopulations. May confound local and metapopulation N_e when migration rate is high.	Do et al. 2014; Hui, Brenas, and Burt 2021; Hui and Burt 2015; R. S. Waples 1989
	Contemporary	Sibship assignment	COLONY 2	Frequencies of full- and half-sib dyads inferred from genotype data of single cohorts	Assumes discrete generations. Not sensitive to spatial genetic structure, but sensitive to the assumption of random sampling. Provides N_b of the sampled area.	Jones and Wang 2010; Wang 2009
	Contemporary	Approximate Bayesian Computation (ABC)	OneSAMP	Summary statistics obtained from genotype data	Computationally intensive when the number of loci is large. Estimates N_{ex} .	Hong et al. 2024; Tallmon et al. 2008

to interpret the N_e value if the population was not isolated across its entire history (Pérez-Sorribes et al. 2024).

To illustrate that there is not a single measure of N_e that provides the definitive effective size of a population, we can consider the human population history. From a modern sample of human genomes, the genome-wide nucleotide diversity reflects that our early ancestors underwent a prolonged bottleneck of about 1000 (effective) individuals (Hu et al. 2023). The long-term coalescent N_e Co, which we can consider as the harmonic mean of the N_e of each generation, is still very low in the human population (tens of thousands) because of that bottleneck. However, if we were to calculate the contemporary N_e Meta of the global human population based on the variance in reproductive success, we would find N_e Meta to be around 3.8 billion. Both estimates of N_e are correct and useful, but they represent different concepts.

To visualize the challenges in estimating N_e in metapopulations, we can consider dynamic metapopulations (spatial variation) across time (temporal variation) to understand that some subpopulations could disappear as time progresses. Figure 1A,C provide schematic representations illustrating the dynamics of metapopulations and the challenges in estimating N_e . Depending on the methodology and sampling design, from a sample taken at t_0 , one could estimate the contemporary N_e of the metapopulation (N_e Meta) or the contemporary local N_e of a single subpopulation (N_e x). But sampling population could also be used to estimate the ancestral ($t-2$) N_e Meta. However, there are also countless ways model assumptions can be violated that can under- or overestimate the particular N_e of interest, especially when N_e 's different spatial and temporal types are confused.

When a N_e exceeds 500–1000 individuals, populations can generally maintain sufficient adaptive genetic variation (Frankham, Bradshaw, and Brook 2014). The 2022 Kunming-Montreal Global Biodiversity Framework (GBF) has included specific goals and targets on safeguarding genetic diversity in the wild (Goals A and C; Targets 4 and 13) and specifically recognized that an effective size larger than 500 is required to maintain evolutionary potential (headline indicator A.4; CBD 2022). This criterion will likely seep through other biodiversity policy and management instruments (e.g., biodiversity strategies and action plans; Hoban, Hvilsum, et al. 2024), hence being of further significance in future conservation planning. This raises the question of both method and N_e type: what N_e should we focus on for present-day conservation questions, and how can we best estimate it?

1.2 | Operationalizing N_e Estimation for End Users: Bridging Science and Conservation Practice for Better Management of Genetic Diversity Within Species

Reflecting the extensive body of work dedicated to developing methods and tools for estimating N_e using simulations and empirical data (e.g., Frankham, Bradshaw, and Brook 2014; Gilbert and Whitlock 2015; Marandel et al. 2020; Nadachowska-Brzyska, Konczal, and Babik 2022; Neel et al. 2013; Nunney 1999, 2016; Palstra and Ruzzante 2011; Ryman, Laikre, and Høssjer 2019;

Tallmon, Luikart, and Beaumont 2004; Waples and Do 2010), to name a few, the members of Working Group 2 of the EU Cost Action G-BIKE; (<https://g-bikegenetics.eu/en>) organized a workshop to focus on the evaluation and implementation of N_e in biodiversity monitoring for better species management. A genetic diversity indicator that can use census population size (N_c) as a proxy for N_e (Hoban et al. 2020; Laikre et al. 2020) was recently adopted by CBD parties as a headline indicator A.4 for the monitoring framework of the Kunming-Montreal GBF. Hoban et al. (2020) defined indicator 1 as “The number of populations within a species with an effective population size (N_e) above 500 compared to the number below 500” (headline indicator A.4 of the GBF, CBD 2022). When no direct estimate of N_e is available, typically due to a lack of genetic data, it is suggested to use N_c as a proxy, using as a rule of thumb an averaged N_e/N_c ratio of 0.10. Although this may be a conservative estimate (Clarke et al. 2024), the $N_e > 500$ threshold has been criticized for being overly liberal in some cases, rather requiring $N_e > 1000$ (Frankham, Bradshaw, and Brook 2014), especially in species with a low fecundity and therefore a high N_e/N_c ratio (Pérez-Pereira et al. 2022). The proxy-based methodology, using largely N_c or proxies of N_c to assess the CBD indicators, was developed elsewhere (Hoban, da Silva, et al. 2023; Mastretta-Yanes et al. 2024; Hoban, da Silva, et al. 2024; Hoban, Hvilsum, et al. 2024).

In our hybrid meeting in Braşov, 26 experts from diverse European and international origins convened to discuss the pragmatic challenges of conservation implementing theoretical frameworks (Figure 2), and this was followed by numerous virtual meetings that included additional experts. Presentations at the workshop from attendees on the projects they were involved in included overviews of possible issues, available data, missing data, prospects, and potential barriers and solutions to N_e indicator estimation in their chosen taxa: Iberian lynx (*Lynx pardinus*), Wolverine (*Gulo gulo*), Grey wolf (*Canis lupus*), Alpine ibex (*Capra ibex*), European beech (*Fagus sylvatica*), maritime pine (*Pinus pinaster*), otters (*Lutra lutra*), and Wels catfish (*Silurus glanis*). We note that the N_e being referred to here is the short term/recent N_e , which determines the maintenance of genetic diversity, not the long-term or coalescent N_e .

The central goal of the workshop was to conduct the groundwork to standardize protocols for the application of N_e estimators, with a focus on (1) determining the most robust methods across various scenarios and (2) elucidating the process of deriving a consensus N_e estimate for species of conservation significance, particularly as it was under the umbrella of EU Cost in the European context. The workshop evaluated widely distributed animal and plant species for which both census population size estimates and independent calibration of N_e estimates were available, laying the foundation for our analyses and evaluation. During the workshop, several challenges emerged in understanding N_e estimates. These included how to delineate populations across the continent and nations and establish the spatial scale at which N_e is to be estimated, how to best select methods based on the interplay between sampling design constraints, life-history traits (social structure, dispersal capacity, overlapping generations, etc.), and methodological assumptions, and how we could gain better knowledge on these examples through forward simulations.



FIGURE 2 | Participants of the international workshop, funded by the COST Action 18134, Genomic Biodiversity Knowledge for Resilient Ecosystems (G-BiKE), August 2022 in Braşov, Romania. Participants who joined virtually are not shown.

Addressing the challenge of defining populations involves balancing management considerations with biological and environmental factors that account for admixture, demographic and spatial expansion, but also ongoing fragmentation and range shifts of natural populations. The challenge of selecting the best tools and estimates involves consideration of temporal design (e.g., single vs. multiple events), sampling designs, and selecting appropriate approaches to estimate specific types of N_e while being aware of the assumptions and biases when assumptions are not met and of the type of N_e that is estimated with each approach.

For the effective size larger than the N_{e500} criterion, the true N_e of interest is the additive variance N_e , which defines the rate of loss of additive genetic variance, or evolutionary potential (Ryman, Laikre, and Hössjer 2019). This is mostly a theoretical concept, but it is best approximated by the contemporary metapopulation (N_{eMeta}) or the realized inbreeding effective size (N_{eRx}) of subpopulations within a metapopulation (Ryman, Laikre, and Hössjer 2019). In isolated populations, N_{eX} and N_{eRx} are evidently identical. In theory, some temporal methods (estimating N_e from the net change in gene diversity across a certain time interval) could estimate N_{eRx} . In practice, this often depends on additional assumptions, and estimates can be hugely biased (Nunney 2016). Also, temporal methods that are based on measuring the variance in allele frequency across time will tend to yield a value close to N_{eX} (Ryman, Laikre, and Hössjer 2019). In practice, we are often limited to methods that estimate N_{eX} , whereas the metric of interest for the N_{e500} threshold value is rather N_{eMeta} . As long as the absolute migration rate is low, the influence of gene flow on contemporary N_{eX} is small. To estimate N_{eX} , two methods are frequently used, and their sensitivities to

assumptions and their performance (precision and accuracy in relation to sample size and the number of markers) have been tested extensively *in silico* (Do et al. 2014; Neel et al. 2013; Wang 2016; Waples 2021; Waples, Antao, and Luikart 2014). The sibship frequency method (Jones and Wang 2010) is not dependent on random mating and is rather insensitive to spatial genetic structure (Wang 2016). It requires random sampling and a good representation across the entire population distribution. However, when the true N_e is very large, it becomes imprecise unless the sample size is $>10\%$ of the true N_e . The accuracy of the estimates obtained with the linkage-disequilibrium method (Waples and Do 2008) is strongly dependent on spatial genetic structure and on the sampling strategy adopted; for instance, the method can provide unbiased estimates when local sampling is carried out in a subpopulation model unless the migration rate is high (Cox, Neyrinck, and Mergeay 2024; Neel et al. 2013). Mergeay et al. (2024) provide examples of this sensitivity compared to the sibship frequency method.

As long as metapopulations consist of well-connected subpopulations (with one migrant per generation as a bare minimum for connectivity), we can, in theory, approximate N_{eMeta} by taking the sum of the N_{eX} of all subpopulations (Cox, Neyrinck, and Mergeay 2024; Mergeay et al. 2024; Ryman, Laikre, and Hössjer 2019). Note that this approach is intended for an island model with symmetrical gene flow. In other cases (e.g., asymmetrical gene flow, very uneven subpopulation sizes, linear metapopulations, populations with extensive two-dimensional isolation by distance, frequent extinction-recolonization dynamics), more targeted models may be needed to estimate N_{eMeta} (Maruyama and Kimura 1980; Whitlock and Barton 1997; Nunney 1999). When gene flow falls below one migrant per

generation, the correlation of allele frequencies among subpopulations becomes very weak, gene diversity within subpopulations rapidly decreases relative to the metapopulation, and inbreeding within subpopulations becomes increasingly important (Wright 1951). Consequently, it is of little use to estimate N_e in a conservation context when gene flow drops below one migrant per generation.

Addressing taxon-specific issues involves identifying mating systems, spatial structuring of populations, geographic distribution within countries, and availability of genetic data. We also tried to understand the relative influences of drift, inbreeding, and selection, which constitutes another major challenge. As such, N_e estimation tools used in conservation need to be forward compatible: we must ensure that they are easily integrated, among other things, with simulation approaches, species distribution modeling and climate change models (Haller and Messer 2019).

Tools such as GONE (Santiago et al. 2020) seem particularly relevant to reconstructing N_e changes that happened since the large-scale influence of humans on biomes, ecosystems, and populations, as they provide a view on N_e in the recent past before we started monitoring biodiversity declines but after we started having a clear impact. Such tools complement methods that estimate the coalescent N_e or reconstruct long-term N_e trajectories (Excoffier et al. 2013; Gutenkunst et al. 2009; Li and Durbin 2011; Liu and Fu 2015; Schiffels and Durbin 2014) and provide helpful insight into pre-anthropogenic reference values of N_e or target values for ecosystem and population restoration in the long run. Even though GONE has been extensively tested

in silico (Novo et al. 2023; Santiago et al. 2020) and even with experimental populations (Novo et al. 2023), empirical testing with real and often messy data, using legacy datasets to explore the benefits and limitations of the method, remains rare and needed (Gargiulo, Decroocq, et al. 2024).

Two complementary approaches were proposed during the workshop (Figure 3): one focusing on constructing hypothetical datasets using simulations to test a range of alternative scenarios on N_e estimation where biases might exist (Figure 3, scenario A). The second approach was concerned with the manipulation of existing empirical datasets to mimic some of the likely biases that may occur and assess the effects of biases on the estimates (Figure 3, scenario B). The aim of these approaches was also to evaluate the performance of different software and how spatial and/or temporal scales affect N_e estimates.

In terms of parameter manipulation discussed during the workshop, our list is not exhaustive and could be modified based on the characteristics of the dataset. It is augmented by Hoban, Bertorelle, and Gaggiotti (2012), where additional information can also be found, including details on simulation applications, evaluation of simulator capabilities, and guidance for their use. Moreover, Hoban (2014) analyzed several case studies illustrating the use of simulations, elucidating their specific advantages and necessity, and exploring alternative or complementary (non-simulation) approaches.

The suggestions raised during the workshop for parameter manipulation included *sample characterization* such as (i) *sample sizes* that might be simply manipulated, for instance,

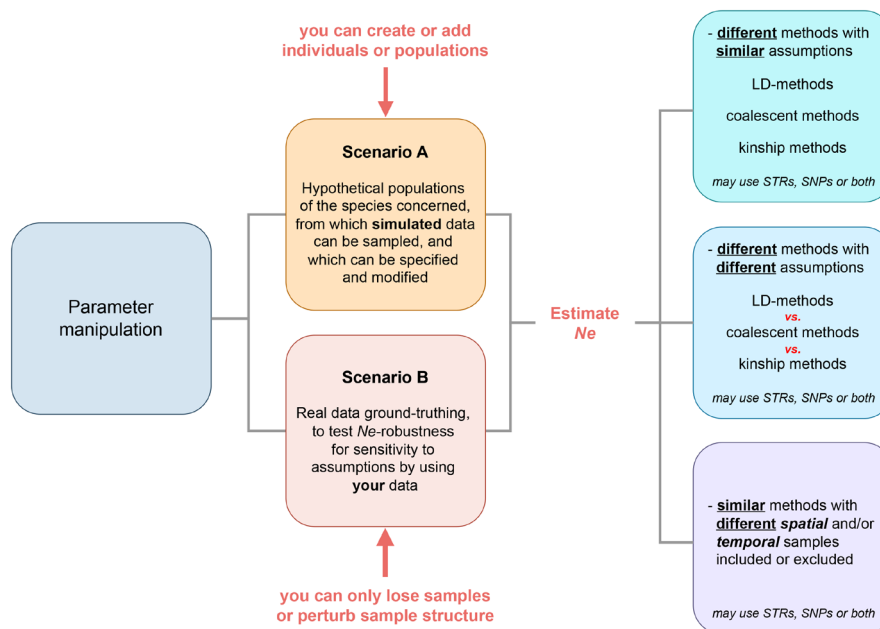


FIGURE 3 | A schematic overview to evaluate various methods used to estimate the N_e by incorporating different techniques and underlying assumptions: The two approaches involved in parameter manipulation during the workshop were constructing hypothetical datasets using simulations to test a range of alternative scenarios on N_e estimation (scenario A) and manipulation of existing empirical datasets to mimic some of the likely biases that may occur (scenario B). Genetic markers such as short tandem repeats (STRs or microsatellites) or single nucleotide polymorphisms (SNPs) are used independently or in combination. N_e estimation involves different methods with divergent assumptions, including LD-based approaches and kinship analysis. Similar to the first scenario (A), these methods may use STR, SNP, or a blend of both. Finally, N_e estimation strategies may involve consistent methodologies with variations in including or excluding spatial and/or temporal samples. These variations allow for an evaluation of how sample selection influences the accuracy of N_e estimation. These analyses may use STR, SNP, or a combination of genetic markers.

by subsampling a reduced number of individuals: the size of the sample should be realistic as a function of N_c , and (ii) *sample distribution*, for example, different relevant sampling designs could be considered, depending on the population configuration and structure, including within meta, continuous, and isolated populations. An important aspect raised was how samples should be distributed based on what is known about dispersal and neighborhood size for the species (e.g., Cox, Neyrinck, and Mergeay 2024). Additional specific elements discussed were the *single temporal* sample estimates that can be compared with *multisample temporal* estimates where temporal sampling is available and, for the latter, different *times between sampling events*, for example, as a function of generation time, can be considered to compare N_e estimates across different timespans. Parameter manipulation requires consideration of *relatedness*; thus, keeping all relatives in the data versus pruning relatives can be done for estimations based on LD to ensure families do not dominate the dataset and *migration* (N_m , gene flow) is testing > 1 or < 1 (by moving genotypes from one population to another).

When DNA or genetic data is unavailable, as in many countries and regions (Pearman et al. 2024), proxy-based indicator values of N_e are extremely important to evaluate and track multiple species affordably (Mastretta-Yanes et al. 2024). Such proxy-based indicators might, for instance, identify populations in urgent need of genetic monitoring and management (e.g., small and/or isolated populations), for which genetic data can be produced for a full assessment of genetic composition and change, such as small and/or isolated populations (Hoban, Paz-Vinas, et al. 2024). A small population (in terms of census size) might be indicative of low effective population sizes, hence compromising the maintenance of genetic diversity over time by these populations. Once such populations are identified using affordable proxy-based indicators, full genetic assessments needing DNA data production could preferentially target such populations to evaluate migration rates, inbreeding, the occurrence of genetic bottlenecks, etc. In this Special Issue of Evolutionary Applications, Mergeay et al. (2024) found a good correspondence between direct N_e estimates and N_c values for wolf populations in Europe. It is important, however, to study more in-depth the relationship between N_e and proxies across species to identify situations where proxies serve their purpose but also where we need actual genetic data (Hoban, da Silva, et al. 2024). This approach can help us focus on species and populations where genetic and genomic resources must be developed.

1.3 | Legacy Datasets to Test Method Suitability, Sampling Designs and Model Assumptions

We searched for so-called legacy datasets: public or own genotypic data archives with sufficient metadata, large sample sizes, and, where possible, genotypic information from different genetic marker types. Especially large and spatially explicit sampling designs would allow us to test the importance of spatial sampling design, sample size, etc., which pertains to model assumptions of particular N_e estimation methods. A legacy dataset is a large, well-annotated archived dataset that allows the calculation of specific properties of real populations, which can also be verified with independent data. In the context of population

genetics, and especially of N_e estimations, these would be large genotypic or genomic datasets of real species and populations with well-known properties associated with metadata of census size, age of individuals, possibly individual-based spatial and ecological information or pedigree data, and data on reproduction and other life history traits (Pierson et al. 2018). Such datasets can be used to test the sensitivity of N_e estimation methods to underlying model assumptions, detect biases, and, by comparison, find what methodology and spatial sampling design best fit the known or expected N_e . It comes closest to simulated data, with the advantage of no dependence on the simulation assumptions. Analyses of such datasets can support the development of guidelines for sampling and analyzing species and populations with similar properties. A disadvantage of legacy datasets is that, unlike simulated data, the entire population is unlikely to be fully sampled, whereas simulated data can provide a complete sample.

In practice, only some legacy datasets fulfill all relevant criteria to carry out sensitivity analyses and test model assumptions. However, there are hundreds, if not thousands, of datasets that actually meet many or most of these criteria (Leigh et al. 2021), and which allow us to test aspects of model assumptions in N_e estimation. To develop standards for monitoring N_e , we sought to explore using legacy datasets to test the sensitivity of molecular N_e estimation methods to underlying model assumptions and optimize sampling designs for future projects. This Special Issue (*Effective population size in conservation and biodiversity monitoring*) presents papers that explore and analyze legacy datasets to better understand the sensitivity of N_e estimation methods to model assumptions (Cox, Neyrinck, and Mergeay 2024; Gargiulo, Decroocq, et al. 2024; Mergeay et al. 2024; Pérez-Sorribes et al. 2024).

2 | Working Group Discussions and Output

2.1 | Challenges to N_e Estimation Differ Among Higher Taxa

Specific working groups discussed factors that affect N_e estimations in species of animals (reptiles, amphibians, fish, and mammals) and plants, and the available datasets were evaluated. The discussions also helped identify the common factors influencing N_e estimates across taxa. The potential influential factors on the estimation of N_e are reflected and summarized in a multi-layer schematic that represents aspects of the available data, species life histories, and population characteristics (Figure 4). Finally, the discussions helped working groups to identify datasets that have served as the basis for research that is reported in this Special Issue and elsewhere.

2.2 | Population Structure: Continuous Distribution, Isolation-By-Distance, and Recent Expansions

Accurate identification of populations and population structure may impact N_e estimation and arise in multiple higher taxa. Recent population recovery has resulted in the re-establishment of connectivity between previously isolated, genetically

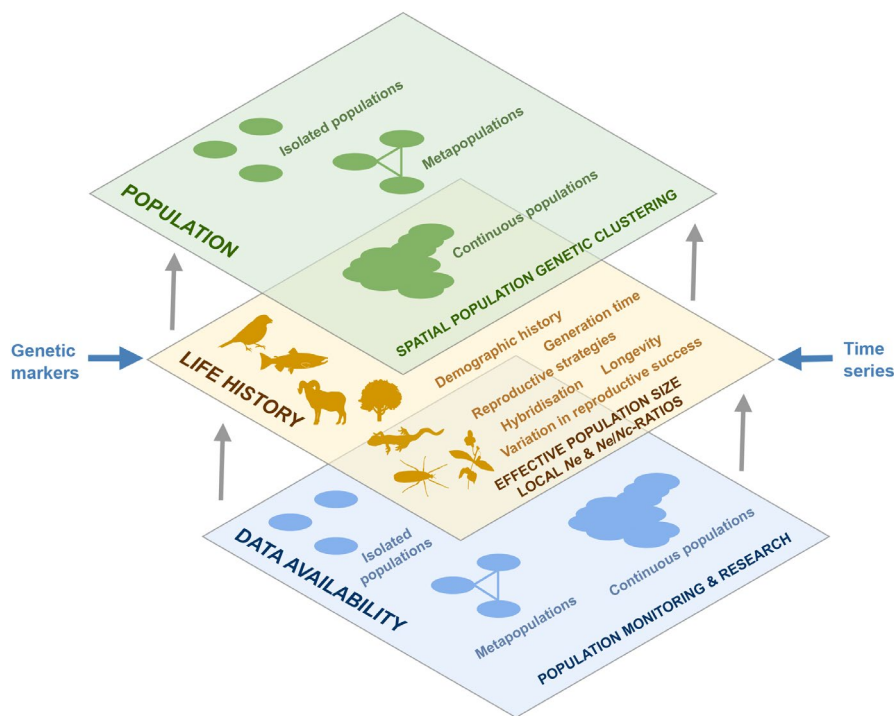


FIGURE 4 | A general representation of the complexity of estimating effective population size (N_e). The figure presented is a conceptual model illustrating the complex framework used to assess genetic indicators in the context of conservation genetics. It details the interactions between the different levels of analysis and the factors that influence the assessment of genetic diversity in various species. At the highest level, ‘spatial genetic clustering of populations’ is highlighted, distinguishing between isolated populations, metapopulations, and continuous populations. This classification is key to understanding gene flow and genetic structure, which are fundamental to conservation strategies. At the middle level, the ‘life history level’ includes aspects such as demographic history, generation time, reproductive strategies, differences in reproductive success, and hybridization. These elements influence the effective population size and the N_e/N_c ratio, which are essential parameters in the study of population genetics; this life-history data primarily provides insights into local N_e . Genetic markers and time-series data are essential for comprehending the structure and dynamics of populations at a spatial scale, yet they have the potential to introduce biases that could diminish the effectiveness of the analysis. The framework underscores the necessity of synthesizing data from genetic markers with an in-depth understanding of species’ life histories. Additionally, it highlights the imperative to secure the availability of extensive genetic data, which is critical to informing effective conservation strategies.

differentiated units; what appears to be a spatially contiguous population may have significant spatial genetic structuring and violate the assumption of a single panmictic population. Some carnivores and larger mammals in Europe have experienced recent population recoveries from small isolated populations yet may still exhibit substantial spatial genetic structuring (Thomas et al. 2022). Challenges emerged from complexity in defining populations for accurate N_e estimation, especially for mammal species exhibiting hybridization (Adavoudi and Pilot 2021) and population structure and continuous species distributions (Adavoudi and Pilot 2021; Iacolina et al. 2021; Randi 2007). Identifying genetic boundaries among populations is important for determining appropriate sampling strategies to avoid pooling populations. In contrast, widely distributed species of plants may experience isolation by distance (IBD) and absence of panmixia, which may be difficult to compensate for with any sampling strategy. Similarly, N_e estimation for fish and other riverine species populations is likely complicated by spatial solid patterns of genetic variation, isolation by distance, and potentially metapopulation dynamics. The influence of population traits on N_e , particularly differences in demographic stochasticity and reproductive variance, requires further investigation (May et al. 2023; Wright, Schofield, and Mathews 2021).

Different characteristics affecting N_e estimation were identified for fish, depending on whether the species is a cartilaginous or bony fish species and the species’ main habitat (marine, rivers, lakes, and ponds). Difficulties in N_e estimation arising from the typically large N_c and N_e and associated stochasticity observed for marine fishes have been discussed previously (e.g., Marandel et al. 2019; Montes et al. 2016). However, cartilaginous fish like rays and sharks, while mainly marine, seem not to display N_e values as high as those of bony marine fishes (Hoban et al. 2020), and the potential effects of the low reproductive output and high longevity of cartilaginous fishes compared to bony marine fishes were discussed. N_e variation within and across taxonomic groups has recently been addressed (Hoban, da Silva, et al. 2024). A talk during the workshop further addressed technical limitations related to estimating N_e and N_c in a large-bodied fish species (*Silurus glanis*) that inhabits large river systems and has undergone a demographic expansion (Paz-Vinas et al. 2024). Finally, the determination of population extent is further complicated by hybridization between species. For instance, for amphibians, the groups identified the potential importance of hybridization following secondary contact (e.g., for salamanders, Bruni et al. 2023; Patton et al. 2020). Hybridization could potentially affect N_e estimation in many plant species.

In contrast, the group discussions recognized population characteristics that may facilitate N_e estimation when population definition or extent is relatively easy to define. The strong spatial structure, genetic isolation, and clear population boundaries of many reptile species and pond-breeding amphibians were highlighted as factors that might facilitate N_e estimation, as these characteristics of populations may approximate the many assumptions made by most N_e estimation methods. In fish species inhabiting isolated lakes and ponds, conformity to some assumptions of the Wright-Fisher model (i.e., no immigration, constant population size) might be more frequently met than in complex riverscapes, limiting potential biases in N_e estimation (Neel et al. 2013; Waples 2024b).

2.3 | Effects of Life History Variation on N_e Estimation

Compared to potential impacts that affect species across several taxa, some population characteristics appear fairly restricted to one taxon. For example, the effects of the existence of plant seed banks on N_e estimation were identified as requiring study. Seed banks result in overlapping generations even in annual species, and models need development to examine the potential effects on N_e estimation. Seed banks can compensate for fluctuations in population sizes typical of some plant species by increasing N_e and delaying the loss of genetic diversity (Nunney 2002). Also, smolts of Pacific salmon may also reproduce at varying ages (Waples 2006). Additional limitations on N_e estimation are associated with life-history traits, including mating system and reproductive strategies, which influence how N_e varies for N_c and the magnitude of the N_e/N_c ratio (Gargiulo, Budde, and Heuertz, 2024). Some subsequent research has attempted to use existing literature and data to predict the direction of the bias associated with these limitations (e.g., Gargiulo et al. 2023; Neel et al. 2013; Waples 2016; Waples et al. 2013). Such predictions are challenging, as the combination of different life-history traits would affect N_e and the N_e/N_c ratios possibly in contrasting directions and magnitudes.

The working groups concurred that overlapping generations, demographic fluctuations, and additional factors in long-lived species certainly complicate N_e estimation. For example, in a study focusing on mammals, Pérez-Sorribes et al. (2024) leveraged two open-access genomic datasets from wolf populations in Minnesota and Scandinavia. These data sets represented populations with differing histories and provided known census population size N_c over the past 40–120 years. High-density SNP genotypes or whole genome sequencing (WGS) data were available for testing how well GONE (Santiago et al. 2020) reconstructed real demographic changes over time, given their complex histories and certain known violations of underlying assumptions. The authors found good concordance between estimated N_e and trends in census size data, but the reconstruction of N_e highlighted the difficulty of interpreting results in spatially structured populations that had undergone demographic fluctuations.

Additional life history characteristics were identified as factors potentially influencing N_e estimation broadly across taxa. The

propensity of some reptile lineages to have morphologically cryptic species, as in some lizards (Pinho et al. 2022), and/or overlapping generations in long-lived species was noted. Analytical challenges are presented by the huge genome sizes of some amphibians (Liedtke et al. 2018) and the strong variation in N_e/N_c ratio reported for some amphibians (Hoban et al. 2020). Polyploidy and genome size could also present analytical challenges in plants. Integrating N_e estimates with other data types such as census population size, demography and population structure, and ecological data is extremely important as it allows the calibration of molecular N_e estimation methods and tests the sensitivity to violations of model assumptions (Mergeay et al. 2024).

2.4 | Aspects of Datasets

The group discussed factors linked to sampling strategies known to bias N_e estimation, opportunities generated by new N_e software for use with plant data, and that very few plant species have readily available genomic resources, such as reference genomes at the chromosome level. Census sizes (N_c) are also mostly unavailable and difficult to determine in plants. This makes some recently developed software, such as GONE (Santiago et al. 2020), unsuitable. In contrast, resources are extensive for some charismatic mammal species (Mergeay et al. 2024; Pérez-Sorribes et al. 2024). Similarly, Gargiulo, Decroocq, et al. (2024) explored the limitations of plant genomic datasets when estimating recent historical N_e using GONE. In particular, genomic datasets from non-model species are usually derived from reduced-representation methods, and linkage maps and reference genomes at the chromosome level are seldom available, all factors that present constraints to using GONE. The authors extracted genomic data from four plant species and showed how the accuracy and precision of N_e estimates changed with the extent of missing data, the number of SNPs and individuals sampled, and the lack of information about the location of SNPs on chromosomes. The latter factor, in particular, had not been previously explored with empirical data and produced a significant upward bias in the N_e estimation. The authors also evaluated the influence of population structure and gene pool admixture for one of the datasets, pointing out that this is influenced by the demographic history of each gene pool (e.g., recent bottlenecks). Furthermore, they evaluated the consistency of the N_e estimates obtained with GONE for the most recent generations and the contemporary N_e estimates (Santiago et al. 2024) and based on NeEstimator (Do et al. 2014). They showed a clear agreement between the estimates obtained with the latter two programs. Finally, they proposed a set of recommendations when estimating N_e in plants using GONE.

2.5 | Immediate Response to Identified Issues

Through the momentum of this workshop, G-BIKE COST Action funded targeted Short Term Scientific Missions (STSMs) of approximately a month to target specific questions in depth, thereby testing the robustness of molecular methods and helping produce guidelines for particular taxa and situations. The STSM on plants explored some of these biological and technical limitations. Results from these scientific missions have

been reported (Cox, Neyrinck, and Mergeay 2024; Gargiulo, Decroocq, et al. 2024; and Pérez-Sorribes et al. 2024).

2.6 | Outstanding Questions to Be Addressed

Building on decades of theoretical and empirical work on N_e , we aimed to prepare the groundwork for testing the sensitivity of assumptions and gaining a deeper understanding of the reliability of N_e estimates for the latest Global Biodiversity Framework. We have presented and discussed the challenges to arrive at the optimal decision. It is crucial that scientists meticulously review all existing methods and tools for N_e , evaluate their current application in various species groups, and ascertain their appropriateness for future management. This comprehensive evaluation ensures that our decision-making process is grounded in the most reliable evidence available, enabling us to make informed choices.

We lack a comprehensive synthesis of practical field applications and technological advancements in conservation genetics and genomics that would serve as the foundation for developing operational guidelines tailored to end users in conservation. Clearly, there is a need for standardized protocols, tools, and resources that can be readily implemented by practitioners. Further, ongoing collaboration between researchers, conservation practitioners, policymakers, and stakeholders is essential to ensure that guidelines are relevant, practical, and effectively disseminated. Fortunately, several studies on this challenge have been published in the past (Heuertz et al. 2023; Hoban, Bruford, et al. 2021; Holderegger et al. 2019; Kershaw et al. 2022; Lundmark et al. 2017; Taft et al. 2020). Lastly, there is a need for capacity building and training for practitioners to use the proposed tools and for scientists to be trained in the practicalities of management and constraints of real-world situations. Fostering collaboration and knowledge exchange between practitioners and scientists could enable the advancement of refined field protocols for sampling and protocols for data analysis. Therefore, also research investment is crucial for advancing sampling protocol optimisation, and refining analytical techniques. It further includes advancements in DNA sequencing technologies, bioinformatics software, and field sampling equipment. Rigorous validation and testing of protocols in real-world conservation scenarios are necessary to ensure their effectiveness and reliability. It involves conducting pilot studies and field trials to evaluate the performance of new protocols across different species and environmental conditions. Such investments and collaboration will enhance training programs and educational resources to equip conservation practitioners and the next generation of scientists with the knowledge and skills. Integrating refined sampling and analysis protocols into broader conservation planning frameworks ensures that genetic data are effectively utilized in decision-making processes.

3 | Conclusions

Despite making foundational progress in exploring current knowledge on methods of N_e estimation, some open questions need further investigation. A primary concern is the lack of a comprehensive synthesis of field applications and technological

advancements, which would be useful as a foundation for guidelines explicitly tailored to end-users interested in N_e estimation and application of Essential Biodiversity Variables (EBVs) for genetic composition (Hoban et al. 2022). Furthermore, there is an urgent need to standardize protocols, tools, and resources by using genomics data that practitioners can readily implement, thereby ensuring consistency, reliability, and comparability in their conservation strategies and population monitoring. Strengthening collaboration among various stakeholders to share expertise, data, and resources is necessary for resolving assumptions and developing strategies and solutions for data analyses. This can consolidate effective population size N_e as a headline indicator for the Global Biodiversity Framework and additionally contribute to practitioners' acceptance of N_e as an EBV (Hoban et al. 2022).

Investment in genetic and genomic approaches as well as tools is pivotal to enhancing the efficiency and accuracy of sampling and analysis. Rigorous validation and testing of new protocols in real-world scenarios are necessary to ascertain their effectiveness and reliability across different species and environmental conditions. The provision of training programs and educational resources is essential to equip the new generation of conservation practitioners and scientists with the necessary skills and knowledge. Nevertheless, we have laid some groundwork and advocated for increased efforts to develop practical and operational guidelines for end-users in the field of conservation genetics and genomics. Further efforts, projects, and initiatives are necessary to better address the current application and reliability of N_e estimates in various species groups and ascertain their appropriateness to provide practical and operational guidelines for conservation genetics and genomics end-users.

Affiliations

¹Department of Wildlife, National Institute for Research and Development in Forestry 'Marin Dracea', Braşov, Romania | ²Department of Silviculture, Faculty of Silviculture and Forest Engineering, Transilvania University of Braşov, Braşov, Romania | ³Research Institute for Nature and Forest, Geraardsbergen, Belgium | ⁴Applied Population Genetics and Conservation Genomics, Department of Biology, KU Leuven, Leuven, Belgium | ⁵Department of Forest Production and Products, University of Ibadan, Ibadan, Nigeria | ⁶Department of Biology, Istiklal Yerleskesi, Budur Mehmet Akif Ersoy University, Science and Art Faculty, Burdur, Türkiye | ⁷Dokuz Eylül University, Buca Education Faculty, Mathematics and Science Education, Biology Education, İzmir, Türkiye | ⁸Department of Evolutionary Biology and Environmental Studies, University of Zurich, Zurich, Switzerland | ⁹Gran Paradiso National Park, Alpine Wildlife Research Center, Noasca, Italy | ¹⁰Research Institute of Wildlife Ecology, University of Veterinary Medicine Vienna, Vienna, Austria | ¹¹University of Primorska, Faculty of Mathematics, Natural Sciences and Information Technologies, Koper, Slovenia | ¹²Faculty of Environmental Protection, Velenje, Slovenia | ¹³Department of Animal Science, University of Zagreb, Faculty of Agriculture, Zagreb, Croatia | ¹⁴Institute of Animal Sciences, Hungarian University of Agriculture and Life Sciences (MATE), Kaposvár, Hungary | ¹⁵Royal Botanic Gardens, Richmond, UK | ¹⁶Department of Ecology and Evolution, Estación Biológica de Doñana, Seville, Spain | ¹⁷INRAE, Univ. Bordeaux, BIOGECO, Cestas, France | ¹⁸WSL Swiss Federal Research Institute, Birmensdorf, Switzerland | ¹⁹The Center for Tree Science, The Morton Arboretum, Lisle, Illinois, USA | ²⁰The Committee on Evolutionary Biology, The University of Chicago,

Chicago, Illinois, USA | ²¹Royal Zoological Society of Scotland, Edinburgh, UK | ²²Institute of Biodiversity and Ecosystem Research at Bulgarian Academy of Sciences, Sofia, Bulgaria | ²³Technical University in Zvolen, Zvolen, Slovakia | ²⁴Czech University of Life Sciences Prague, Faculty of Forestry and Wood Sciences, Department of Forest Ecology, Suchdol, Praha, Czech Republic | ²⁵Department of Zoology, Stockholm University, Stockholm, Sweden | ²⁶Behavioral Ecology Research Group, Leipzig University, Leipzig, Germany | ²⁷Max-Planck Institute for Evolutionary Anthropology, Department of Human Behaviour, Ecology and Culture Deutscher Platz 6, Leipzig, Germany | ²⁸Universite Claude Bernard Lyon 1, Villeurbanne, France | ²⁹Department of Plant Biology and Ecology, Faculty of Sciences and Technology, University of the Basque Country UPV/EHU, Leioa, Spain | ³⁰IKERBASQUE Basque Foundation for Science, Bilbao, Spain | ³¹BC3 Basque Center for Climate Change, Leioa, Spain | ³²Israel Oceanographic and Limnological Research, National Institute of Oceanography, Haifa, Israel | ³³School of Biosciences, Cardiff University, Cardiff, UK | ³⁴Slovenian Forestry Institute, Ljubljana, Slovenia | ³⁵Senckenberg Biodiversity and Climate Research Centre, Frankfurt Am Main, Frankfurt, Germany | ³⁶Norwegian Institute for Nature Research, Trondheim, Norway

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Conflicts of Interest

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Data Availability Statement

Data sharing is not applicable to this article as no datasets were generated or analyzed during the study at hand. *Code Availability*: No code is available to this article as no new data were analyzed or new code was created in this study.

References

- Adavoudi, R., and M. Pilot. 2021. "Consequences of Hybridization in Mammals: A Systematic Review." *Genes* 13, no. 50: 1–26. <https://doi.org/10.3390/genes13010050>.
- Allendorf, F. W. 2017. "Genetics and the Conservation of Natural Populations: Allozymes to Genomes." *Molecular Ecology* 26: 420–430. <https://doi.org/10.1111/mec.13948>.
- Balloux, F. 2004. "Heterozygote Excess in Small Populations and the Heterozygote-Excess Effective Population Size." *Evolution; International Journal of Organic Evolution* 58: 1891–1900. <https://doi.org/10.1111/j.0014-3820.2004.tb00477.x>.
- Barbato, M., P. Orozco-terWengel, M. Tapio, and M. W. Bruford. 2015. "SNEP: A Tool to Estimate Trends in Recent Effective Population Size Trajectories Using Genome-Wide SNP Data." *Frontiers in Genetics* 6, no. 109: 1–6. <https://doi.org/10.3389/fgene.2015.00109>.
- Bertola, L. D., L. Quinn, K. Hanghøj, et al. 2024. "Giraffe Lineages Are Shaped by Major Ancient Admixture Events." *Current Biology: CB* 34, no. 7: 1576–1586.e5. <https://doi.org/10.1016/j.cub.2024.02.051>.
- Bruni, G., A. Chiochio, G. Nascetti, and R. Cimmaruta. 2023. "Different Patterns of Introgression in a Three Species Hybrid Zone Among European Cave Salamanders." *Ecology and Evolution* 13, no. 8: e10437. <https://doi.org/10.1002/ece3.10437>.
- CBD. 2022. "Kunming-Montreal Global Biodiversity Framework." <https://www.cbd.int/doc/decisions/cop-15/cop-15-dec-04-en.pdf>.
- Chikhi, L., V. C. Sousa, P. Luisi, B. Goossens, and M. A. Beaumont. 2010. "The Confounding Effects of Population Structure, Genetic Diversity and the Sampling Scheme on the Detection and Quantification of Population Size Changes." *Genetics* 186, no. 3: 983–995. <https://doi.org/10.1534/genetics.110.118661>.
- Clarke, S. H., E. R. Lawrence, J.-M. Matte, et al. 2024. "Global Assessment of Effective Population Sizes: Consistent Taxonomic Differences in Meeting the 50/500 Rule." *Molecular Ecology* 33: e17353. <https://doi.org/10.1111/mec.17353>.
- Cox, K., S. Neyrinck, and J. Mergeay. 2024. "Dealing With Assumptions and Sampling Bias in the Estimation of Effective Population Size: A Case Study in an Amphibian Population." *Evolutionary Applications* 17, no. 9: e70015. <https://doi.org/10.1111/eva.70015>.
- Do, C., R. S. Waples, D. Peel, G. M. Macbeth, B. J. Tillett, and J. R. Ovenden. 2014. "NeEstimator v2: Re-Implementation of Software for the Estimation of Contemporary Effective Population Size (Ne) From

- Genetic Data." *Molecular Ecology Resources* 14, no. 1: 209–214. <https://doi.org/10.1111/1755-0998.12157>.
- Drummond, A. J., A. Rambaut, B. Shapiro, and O. G. Pybus. 2005. "Bayesian Coalescent Inference of Past Population Dynamics From Molecular Sequences." *Molecular Biology and Evolution* 22, no. 5: 1185–1192. <https://doi.org/10.1093/molbev/msi103>.
- Excoffier, L., I. Dupanloup, E. Huerta-Sánchez, V. C. Sousa, and M. Foll. 2013. "Robust Demographic Inference From Genomic and SNP Data." *PLoS Genetics* 9: e1003905. <https://doi.org/10.1371/journal.pgen.1003905>.
- Excoffier, L., N. Marchi, D. A. Marques, R. Matthey-Doret, A. Gouy, and V. C. Sousa. 2021. "fastsimcoal2: Demographic Inference Under Complex Evolutionary Scenarios." *Bioinformatics* 37: 4882–4885. <https://doi.org/10.1093/bioinformatics/btab468>.
- Fady, B., and M. Bozzano. 2021. "Effective Population Size Does Not Make a Practical Indicator of Genetic Diversity in Forest Trees." *Biological Conservation* 253: 108904. <https://doi.org/10.1016/j.biocon.2020.108904>.
- Frankham, R., C. J. A. Bradshaw, and B. W. Brook. 2014. "Genetics in Conservation Management: Revised Recommendations for the 50/500 Rules, Red List Criteria and Population Viability Analyses." *Biological Conservation* 170: 56–63. <https://doi.org/10.1016/j.biocon.2013.12.036>.
- Gargiulo, R., K. Budde, and M. Heuert. 2024. "Mind the Lag: Understanding Delayed Genetic Erosion." *EcoEvoRxiv*: 1–15. <https://doi.org/10.32942/x26s4x>.
- Gargiulo, R., V. Decroocq, S. C. González-Martínez, et al. 2024. "Estimation of Contemporary Effective Population Size in Plant Populations: Limitations of Genomic Datasets." *Evolutionary Applications* 17, no. 5: 1–16. <https://doi.org/10.1111/eva.13691>.
- Gargiulo, R., R. S. Waples, A. K. Grow, et al. 2023. "Effective Population Size in a Partially Clonal Plant Is Not Predicted by the Number of Genetic Individuals." *Evolutionary Applications* 16: 750–766. <https://doi.org/10.1111/eva.13535>.
- Gilbert, K. J., and M. C. Whitlock. 2015. "Evaluating Methods for Estimating Local Effective Population Size With and Without Migration." *Evolution; International Journal of Organic Evolution* 69: 2154–2166. <https://doi.org/10.1111/evo.12713>.
- Goossens, B., and P. Orozco-Ter Wengel. 2023. "Michael (Mike) William Bruford (6th June 1963–13th April 2023)." *Conservation Genetics* 24: 933–934. <https://doi.org/10.1007/s10592-023-01583-6>.
- Gutenkunst, R. N., R. D. Hernandez, S. H. Williamson, and C. D. Bustamante. 2009. "Inferring the Joint Demographic History of Multiple Populations From Multidimensional SNP Frequency Data." *PLoS Genetics* 5: e1000695. <https://doi.org/10.1371/journal.pgen.1000695>.
- Haller, B. C., and P. W. Messer. 2019. "SLiM 3: Forward Genetic Simulations Beyond the Wright-Fisher Model." *Molecular Biology and Evolution* 36: 632–637. <https://doi.org/10.1093/molbev/msy228>.
- Heuert, M., S. B. Carvalho, J. Galindo, et al. 2023. "The Application Gap: Genomics for Biodiversity and Ecosystem Service Management." *Biological Conservation* 278: 109883. <https://doi.org/10.1016/j.biocon.2022.109883>.
- Ho, S. Y. W., and B. Shapiro. 2011. "Skyline-Plot Methods for Estimating Demographic History From Nucleotide Sequences." *Molecular Ecology Resources* 11: 423–434. <https://doi.org/10.1111/j.1755-0998.2011.02988.x>.
- Hoban, S. 2014. "An Overview of the Utility of Population Simulation Software in Molecular Ecology." *Molecular Ecology* 23: 2383–2401. <https://doi.org/10.1111/mec.12741>.
- Hoban, S., F. I. Archer, L. D. Bertola, et al. 2022. "Global Genetic Diversity Status and Trends: Towards a Suite of Essential Biodiversity Variables (EBVs) for Genetic Composition." *Biological Reviews of the Cambridge Philosophical Society* 97: 1511–1538. <https://doi.org/10.1111/brv.12852>.
- Hoban, S., G. Bertorelle, and O. E. Gaggiotti. 2012. "Computer Simulations: Tools for Population and Evolutionary Genetics." *Nature Reviews. Genetics* 13: 110–122. <https://doi.org/10.1038/nrg3130>.
- Hoban, S., M. Bruford, J. D'Urban Jackson, et al. 2020. "Genetic Diversity Targets and Indicators in the CBD Post-2020 Global Biodiversity Framework Must Be Improved." *Biological Conservation* 248: 108654. <https://doi.org/10.1016/j.biocon.2020.108654>.
- Hoban, S., M. W. Bruford, W. C. Funk, et al. 2021. "Global Commitments to Conserving and Monitoring Genetic Diversity Are Now Necessary and Feasible." *Bioscience* 71: 964–976. <https://doi.org/10.1093/biosci/biab054>.
- Hoban, S., J. M. da Silva, A. Hughes, et al. 2024. "Too Simple, Too Complex, or Just Right? Advantages, Challenges, and Guidance for Indicators of Genetic Diversity." *Bioscience* 74: 269–280. <https://doi.org/10.1093/biosci/biae006>.
- Hoban, S., J. M. da Silva, A. Mastretta-Yanes, et al. 2023. "Monitoring Status and Trends in Genetic Diversity for the Convention on Biological Diversity: An Ongoing Assessment of Genetic Indicators in Nine Countries." *Conservation Letters* 16, no. 3: 1–12. <https://doi.org/10.1111/conl.12953>.
- Hoban, S., C. Hvilson, A. Abdeldjalil, et al. 2024. "How can Biodiversity Strategy and Action Plans Incorporate Genetic Diversity Concerns, Plans, Policies, Capacity, and Commitments?"
- Hoban, S., I. Paz-Vinas, S. Aitken, et al. 2021. "Effective Population Size Remains a Suitable, Pragmatic Indicator of Genetic Diversity for all Species, Including Forest Trees." *Biological Conservation* 253: 108906. <https://doi.org/10.1016/j.biocon.2020.108906>.
- Hoban, S., I. Paz-Vinas, R. E. Shaw, et al. 2024. "DNA-Based Studies and Genetic Diversity Indicator Assessments Are Complementary Approaches to Conserving Evolutionary Potential." *Conservation Genetics* 25: 1147–1153. <https://doi.org/10.1007/s10592-024-01632-8>.
- Hoban, S., G. Segelbacher, C. Vernesi, and I. R. Russo. 2023. "Michael W. Bruford (1963–2023)." *Nature Ecology & Evolution* 7: 1349–1350. <https://doi.org/10.1038/s41559-023-02110-z>.
- Holderegger, R., N. Balkenhol, J. Bolliger, et al. 2019. "Conservation Genetics: Linking Science With Practice." *Molecular Ecology* 28: 3848–3856. <https://doi.org/10.1111/mec.15202>.
- Hollenbeck, C. M., D. S. Portnoy, and J. R. Gold. 2016. "A Method for Detecting Recent Changes in Contemporary Effective Population Size From Linkage Disequilibrium at Linked and Unlinked Loci." *Heredity* 117: 207–216. <https://doi.org/10.1038/hdy.2016.30>.
- Hong, A., R. G. Cheek, S. N. De Silva, et al. 2024. "ONeSAMP 3.0: Estimation of Effective Population Size via SNP Data from One Population." *G3 (Bethesda, Md.)* 14: jkae153. <https://doi.org/10.1093/g3journal/jkae153>.
- Hu, W., Z. Hao, P. Du, et al. 2023. "Genomic Inference of a Severe Human Bottleneck During the Early to Middle Pleistocene Transition." *Science* 381: 979–984. <https://doi.org/10.1126/science.abq7487>.
- Hui, T.-Y. J., J. H. Brenas, and A. Burt. 2021. "Contemporary Ne Estimation Using Temporally Spaced Data With Linked Loci." *Molecular Ecology Resources* 21: 2221–2230. <https://doi.org/10.1111/1755-0998.13412>.
- Hui, T.-Y. J., and A. Burt. 2015. "Estimating Effective Population Size From Temporally Spaced Samples With a Novel, Efficient Maximum-Likelihood Algorithm." *Genetics* 200: 285–293. <https://doi.org/10.1534/genetics.115.174904>.
- Iacolina, L., E. Buzan, T. Safner, et al. 2021. "A Mother's Story, Mitogenome Relationships in the Genus *Rupicapra*." *Animals* 11: 1065. <https://doi.org/10.3390/ani11041065>.

- Jones, O. R., and J. Wang. 2010. "COLONY: A Program for Parentage and Sibship Inference From Multilocus Genotype Data." *Molecular Ecology Resources* 10, no. 3: 551–555. <https://doi.org/10.1111/j.1755-0998.2009.02787.x>.
- Kershaw, F., M. W. Bruford, W. C. Funk, et al. 2022. "The Coalition for Conservation Genetics: Working Across Organizations to Build Capacity and Achieve Change in Policy and Practice." *Conservation Science and Practice* 4: 1–14. <https://doi.org/10.1111/csp2.12635>.
- Laikre, L., S. Hoban, M. W. Bruford, et al. 2020. "Post-2020 Goals Overlook Genetic Diversity." *Science* 367: 1083–1085. <https://doi.org/10.1126/science.abb2748>.
- Leigh, D. M., C. B. van Rees, K. L. Millette, et al. 2021. "Opportunities and Challenges of Macrogenetic Studies." *Nature Reviews. Genetics* 22, no. 12: 791–807. <https://doi.org/10.1038/s41576-021-00394-0>.
- Li, H., and R. Durbin. 2011. "Inference of Human Population History From Individual Whole-Genome Sequences." *Nature* 475: 493–496. <https://doi.org/10.1038/nature10231>.
- Liedtke, H. C., D. J. Gower, M. Wilkinson, and I. Gomez-Mestre. 2018. "Macroevolutionary Shift in the Size of Amphibian Genomes and the Role of Life History and Climate." *Nature Ecology & Evolution* 2: 1792–1799. <https://doi.org/10.1038/s41559-018-0674-4>.
- Liu, X., and Y.-X. Fu. 2015. "Exploring Population Size Changes Using SNP Frequency Spectra." *Nature Genetics* 47: 555–559. <https://doi.org/10.1038/ng.3254>.
- Liu, X., and Y.-X. Fu. 2020. "Stairway Plot 2: Demographic History Inference With Folded SNP Frequency Spectra." *Genome Biology* 21, no. 1: 280. <https://doi.org/10.1186/s13059-020-02196-9>.
- Lundmark, C., K. Andersson, A. Sandström, and L. Laikre. 2017. "Effectiveness of Short-Term Knowledge Communication on Baltic Sea Marine Genetic Biodiversity to Public Managers." *Regional Environmental Change* 17, no. 3: 841–849. <https://doi.org/10.1007/s10113-016-1077-1>.
- Marandel, F., G. Charrier, J.-B. Lamy, S. Le Cam, P. Lorange, and V. M. Trenkel. 2020. "Estimating Effective Population Size Using RADseq: Effects of SNP Selection and Sample Size." *Ecology and Evolution* 10, no. 7: 1929–1937. <https://doi.org/10.1002/ece3.6016>.
- Marandel, F., P. Lorange, O. Berthelé, V. M. Trenkel, R. S. Waples, and J.-B. Lamy. 2019. "Estimating Effective Population Size of Large Marine Populations, Is It Feasible?" *Fish and Fisheries* 20, no. 1: 189–198. <https://doi.org/10.1111/faf.12338>.
- Maruyama, T., and M. Kimura. 1980. "Genetic Variability and Effective Population Size When Local Extinction and Recolonization of Subpopulations Are Frequent." *Proceedings of the National Academy of Sciences* 77: 6710–6714.
- Mastretta-Yanes, A., J. M. da Silva, C. E. Grueber, et al. 2024. "Multinational Evaluation of Genetic Diversity Indicators for the Kunming-Montreal Global Biodiversity Framework." *Ecology Letters* 27, no. 7: e14461. <https://doi.org/10.1111/ele.14461>.
- May, S. A., J. J. Hard, M. J. Ford, K. A. Naish, and E. J. Ward. 2023. "Assortative Mating for Reproductive Timing Affects Population Recruitment and Resilience in a Quantitative Genetic Model." *Evolutionary Applications* 16, no. 3: 657–672. <https://doi.org/10.1111/eva.13524>.
- Mergeay, J., S. Smet, S. Collet, et al. 2024. "Estimating the Effective Size of European Wolf Populations." *Evolutionary Applications* 17, no. 12: e70021. <https://doi.org/10.1111/eva.70021>.
- Montes, I., M. Iriondo, C. Manzano, et al. 2016. "No Loss of Genetic Diversity in the Exploited and Recently Collapsed Population of Bay of Biscay Anchovy (*Engraulis encrasicolus*, L.)." *Marine Biology* 163, no. 5: 1–10. <https://doi.org/10.1007/s00227-016-2866-2>.
- Nadachowska-Brzyska, K., M. Konczal, and W. Babik. 2022. "Navigating the Temporal Continuum of Effective Population Size." *Methods in Ecology and Evolution / British Ecological Society* 13, no. 1: 22–41. <https://doi.org/10.1111/2041-210x.13740>.
- Neel, M. C., K. McKelvey, N. Ryman, et al. 2013. "Estimation of Effective Population Size in Continuously Distributed Populations: There Goes the Neighborhood." *Heredity* 111, no. 3: 189–199. <https://doi.org/10.1038/hdy.2013.37>.
- Novo, I., N. Pérez-Pereira, E. Santiago, H. Quesada, and A. Caballero. 2023. "An Empirical Test of the Estimation of Historical Effective Population Size Using *Drosophila melanogaster*." *Molecular Ecology Resources* 23, no. 7: 1632–1640. <https://doi.org/10.1111/1755-0998.13837>.
- Nunney, L. 1999. "The Effective Size of a Hierarchically Structured Population." *Evolution; International Journal of Organic Evolution* 53, no. 1: 1. <https://doi.org/10.2307/2640915>.
- Nunney, L. 2002. "The Effective Size of Annual Plant Populations: the Interaction of a Seed Bank With Fluctuating Population Size in Maintaining Genetic Variation." *American Naturalist* 160, no. 2: 195–204. <https://doi.org/10.1086/341017>.
- Nunney, L. 2016. "The Effect of Neighborhood Size on Effective Population Size in Theory and in Practice." *Heredity* 117, no. 4: 224–232. <https://doi.org/10.1038/hdy.2016.76>.
- Orozco-terWengel, P., E. Coissac, O. Hanotte, F. Pompanon, P. Sunnucks, and P. Taberlet. 2023. "Mike Bruford (1963–2023)." *Molecular Ecology* 3: 5448–5449. <https://doi.org/10.1111/mec.17133>.
- Palstra, F. P., and D. E. Ruzzante. 2011. "Demographic and Genetic Factors Shaping Contemporary Metapopulation Effective Size and Its Empirical Estimation in Salmonid Fish." *Heredity* 107, no. 5: 444–455. <https://doi.org/10.1038/hdy.2011.31>.
- Patton, A. H., M. J. Margres, B. Epstein, J. Eastman, L. J. Harmon, and A. Storfer. 2020. "Hybridizing Salamanders Experience Accelerated Diversification." *Scientific Reports* 10, no. 1: 6566. <https://doi.org/10.1038/s41598-020-63378-w>.
- Paz-Vinas, I., G. Loot, S. Boulêtreau, et al. 2024. "Genetic-Based Inference of Densities, Effective and Census Sizes of Expanding Riverine Meta-Populations of an Invasive Large-Bodied Freshwater Fish (*Silurus glanis* L.)." *bioRxiv* (p. 2024.04.05.588309). <https://doi.org/10.1101/2024.04.05.588309>.
- Pearman, P. B., O. Broennimann, T. Aavik, et al. 2024. "Monitoring of species' Genetic Diversity in Europe Varies Greatly and Overlooks Potential Climate Change Impacts." *Nature Ecology & Evolution* 8, no. 2: 267–281. <https://doi.org/10.1038/s41559-023-02260-0>.
- Pérez-Pereira, N., J. Wang, H. Quesada, and A. Caballero. 2022. "Prediction of the Minimum Effective Size of a Population Viable in the Long Term." *Biodiversity and Conservation* 31: 2763–2780. <https://doi.org/10.1007/s10531-022-02456-z>.
- Pérez-Sorribes, L., P. Villar-Yanez, L. Smeds, and J. Mergeay. 2024. "Comparing genetic Ne reconstructions over time with long-time wolf monitoring data in two populations." *Evolutionary Applications* 12: e70022. <https://doi.org/10.1111/eva.70022>.
- Pierson, J. C., T. A. Graves, S. C. Banks, K. C. Kendall, and D. B. Lindenmayer. 2018. "Relationship Between Effective and Demographic Population Size in Continuously Distributed Populations." *Evolutionary Applications* 11, no. 7: 1162–1175. <https://doi.org/10.1111/eva.12636>.
- Pinho, C., A. Kaliontzopoulou, C. A. Ferreira, and J. Gama. 2022. "Identification of Morphologically Cryptic Species With Computer Vision Models: Wall Lizards (Squamata: Lacertidae: Podarcis) as a Case Study." *Zoological Journal of the Linnean Society* 198, no. 1: 184–201. <https://doi.org/10.1093/zoolinnean/zlac087>.
- Randi, E. 2007. "Phylogeography of South European Mammals." In *Phylogeography of Southern European Refugia: Evolutionary Perspectives on the Origins and Conservation of European Biodiversity*, edited by S. Weiss and N. Ferrand, 101–126. Dordrecht, Netherlands: Springer. https://doi.org/10.1007/1-4020-4904-8_3.

- Ryman, N., L. Laikre, and O. Hössjer. 2019. "Do Estimates of Contemporary Effective Population Size Tell Us What We Want to Know?" *Molecular Ecology* 28, no. 8: 1904–1918. <https://doi.org/10.1111/mec.15027>.
- Ryman, N., L. Laikre, and O. Hössjer. 2023. "Variance Effective Population Size Is Affected by Census Size in Sub-Structured Populations." *Molecular Ecology Resources* 23, no. 6: 1334–1347. <https://doi.org/10.1111/1755-0998.13804>.
- Santiago, E., A. Caballero, C. Köpke, and I. Novo. 2024. "Estimation of the Contemporary Effective Population Size From SNP Data While Accounting for Mating Structure." *Molecular Ecology Resources* 24, no. 1: e13890. <https://doi.org/10.1111/1755-0998.13890>.
- Santiago, E., I. Novo, A. F. Pardiñas, M. Saura, J. Wang, and A. Caballero. 2020. "Recent Demographic History Inferred by High-Resolution Analysis of Linkage Disequilibrium." *Molecular Biology and Evolution* 37, no. 12: 3642–3653. <https://doi.org/10.1093/molbev/msaa169>.
- Schiffels, S., and R. Durbin. 2014. "Inferring Human Population Size and Separation History From Multiple Genome Sequences." *Nature Genetics* 46, no. 8: 919–925. <https://doi.org/10.1038/ng.3015>.
- Schiffels, S., and K. Wang. 2020. "MSMC and MSMC2: The Multiple Sequentially Markovian Coalescent." *Methods in Molecular Biology* 2090: 147–166. https://doi.org/10.1007/978-1-0716-0199-0_7.
- Taft, H. R., D. N. McCoskey, J. M. Miller, et al. 2020. "Research-Management Partnerships: An Opportunity to Integrate Genetics in Conservation Actions." *Conservation Science and Practice* 2, no. 9: 1–8. <https://doi.org/10.1111/csp2.218>.
- Tallmon, D. A., A. Koyuk, G. Luikart, and M. A. Beaumont. 2008. "COMPUTER PROGRAMS: Onesamp: A Program to Estimate Effective Population Size Using Approximate Bayesian Computation." *Molecular Ecology Resources* 8, no. 2: 299–301. <https://doi.org/10.1111/j.1471-8286.2007.01997.x>.
- Tallmon, D. A., G. Luikart, and M. A. Beaumont. 2004. "Comparative Evaluation of a New Effective Population Size Estimator Based on Approximate Bayesian Computation." *Genetics* 167, no. 2: 977–988. <https://doi.org/10.1534/genetics.103.026146>.
- Tenesa, A., P. Navarro, B. J. Hayes, et al. 2007. "Recent Human Effective Population Size Estimated From Linkage Disequilibrium." *Genome Research* 17, no. 4: 520–526. <https://doi.org/10.1101/gr.6023607>.
- Theissinger, K., C. Fernandes, G. Formenti, et al. 2023. "How Genomics Can Help Biodiversity Conservation." *Trends in Genetics: TIG* 39, no. 7: 545–559. <https://doi.org/10.1016/j.tig.2023.01.005>.
- Thomas, N. E., F. Hailer, M. W. Bruford, and E. A. Chadwick. 2022. "Country-Wide Genetic Monitoring Over 21 Years Reveals Lag in Genetic Recovery Despite Spatial Connectivity in an Expanding Carnivore (Eurasian Otter, *Lutra lutra*) Population." *Evolutionary Applications* 15, no. 12: 2125–2141. <https://doi.org/10.1111/eva.13505>.
- Wang, J. 2009. "A New Method for Estimating Effective Population Sizes From a Single Sample of Multilocus Genotypes." *Molecular Ecology* 18, no. 10: 2148–2164. <https://doi.org/10.1111/j.1365-294X.2009.04175.x>.
- Wang, J. 2016. "A Comparison of Single-Sample Estimators of Effective Population Sizes From Genetic Marker Data." *Molecular Ecology* 25, no. 19: 4692–4711. <https://doi.org/10.1111/mec.13725>.
- Waples, R. 2024a. "The idiot's Guide to Effective Population Size." *Authorea Inc*: 1–51. <https://doi.org/10.22541/au.172536694.46670112/v1>.
- Waples, R. S. 1989. "A Generalized Approach for Estimating Effective Population Size From Temporal Changes in Allele Frequency." *Genetics* 121, no. 2: 379–391. <https://doi.org/10.1093/genetics/121.2.379>.
- Waples, R. S. 2005. "Genetic Estimates of Contemporary Effective Population Size: To What Time Periods Do the Estimates Apply?" *Molecular Ecology* 14, no. 11: 3335–3352. <https://doi.org/10.1111/j.1365-294X.2005.02673.x>.
- Waples, R. S. 2006. "Seed Banks, Salmon, and Sleeping Genes: Effective Population Size in Semelparous, Age-Structured Species With Fluctuating Abundance." *American Naturalist* 167, no. 1: 118–135.
- Waples, R. S. 2010. "Spatial-Temporal Stratifications in Natural Populations and How They Affect Understanding and Estimation of Effective Population Size." *Molecular Ecology Resources* 10, no. 5: 785–796. <https://doi.org/10.1111/j.1755-0998.2010.02876.x>.
- Waples, R. S. 2016. "Life-History Traits and Effective Population Size in Species With Overlapping Generations Revisited: the Importance of Adult Mortality." *Heredity* 117, no. 4: 241–250. <https://doi.org/10.1038/hdy.2016.29>.
- Waples, R. S. 2021. "Relative Precision of the Sibship and LD Methods for Estimating Effective Population Size With Genomics-Scale Datasets." *Journal of Heredity* 112, no. 6: 535–539. <https://doi.org/10.1093/jhered/esab042>.
- Waples, R. S. 2022. "What Is Ne, Anyway?" *Journal of Heredity* 113, no. 4: 371–379. <https://academic.oup.com/jhered/article-abstract/113/4/371/6582684>.
- Waples, R. S. 2024b. "Practical Application of the Linkage Disequilibrium Method for Estimating Contemporary Effective Population Size: A Review." *Molecular Ecology Resources* 24, no. 1: e13879. <https://doi.org/10.1111/1755-0998.13879>.
- Waples, R. S., T. Antao, and G. Luikart. 2014. "Effects of Overlapping Generations on Linkage Disequilibrium Estimates of Effective Population Size." *Genetics* 197, no. 2: 769–780. <https://doi.org/10.1534/genetics.114.164822>.
- Waples, R. S., and C. Do. 2008. "Ldne: A Program for Estimating Effective Population Size From Data on Linkage Disequilibrium." *Molecular Ecology Resources* 8, no. 4: 753–756. <https://doi.org/10.1111/j.1755-0998.2007.02061.x>.
- Waples, R. S., and C. Do. 2010. "Linkage Disequilibrium Estimates of Contemporary Ne Using Highly Variable Genetic Markers: A Largely Untapped Resource for Applied Conservation and Evolution." *Evolutionary Applications* 3, no. 3: 244–262. <https://doi.org/10.1111/j.1752-4571.2009.00104.x>.
- Waples, R. S., G. Luikart, J. R. Faulkner, and D. A. Tallmon. 2013. "Simple Life-History Traits Explain Key Effective Population Size Ratios Across Diverse Taxa." *Proceedings. Biological Sciences/The Royal Society* 280, no. 1768: 20131339. <https://doi.org/10.1098/rspb.2013.1339>.
- Whitlock, M. C., and N. H. Barton. 1997. "The Effective Size of a Subdivided Population." *Genetics* 146: 427–441.
- Wright, P. G. R., H. Schofield, and F. Mathews. 2021. "Can Effective Population Size Estimates be Used to Monitor Population Trends of Woodland Bats? A Case Study of *Myotis bechsteinii*." *Ecology and Evolution* 11, no. 5: 2015–2023. <https://doi.org/10.1002/ece3.7143>.
- Wright, S. 1931. "Evolution in Mendelian Populations." *Genetics* 16, no. 2: 97–159. <https://doi.org/10.1093/genetics/16.2.97>.
- Wright, S. 1951. "The Genetical Structure of Populations." *Annals of Eugenics* 15, no. 4: 323–354. <https://doi.org/10.1111/j.1469-1809.1949.tb02451.x>.