



# SERPING1 Variants and C1-INH Biological Function: A Close Relationship With C1-INH-HAE

Christian Drouet<sup>1,2\*</sup>, Alberto López-Lera<sup>3</sup>, Arije Ghannam<sup>4</sup>, Margarita López-Trascasa<sup>5</sup>, Sven Cichon<sup>6,7</sup>, Denise Ponard<sup>8</sup>, Faidra Parsopoulou<sup>9</sup>, Hana Grombirikova<sup>10</sup>, Tomáš Freiburger<sup>10</sup>, Matija Rijavec<sup>11</sup>, Camila L. Veronez<sup>12</sup>, João Bosco Pesquero<sup>12</sup> and Anastasios E. Germenis<sup>9,13</sup>

<sup>1</sup> Department of Infection, Immunity and Inflammation, Institut Cochin, INSERM UMR1016, Université de Paris, Paris, France, <sup>2</sup> Univ. Grenoble-Alpes & Centre Hospitalier Universitaire de Grenoble, Grenoble, France, <sup>3</sup> Hospital La Paz Institute for Health Research (IdiPAZ), CIBERER U-754, Madrid, Spain, <sup>4</sup> KininX SAS, Grenoble, France, <sup>5</sup> Hospital La Paz Institute for Health Research (IdiPAZ), Universidad Autónoma de Madrid, Madrid, Spain, <sup>6</sup> Human Genomics Research Group, Department of Biomedicine, University of Basel, Basel, Switzerland, <sup>7</sup> Institute of Medical Genetics and Pathology, University Hospital Basel, Basel, Switzerland, <sup>8</sup> Centre Hospitalier Universitaire de Grenoble, Grenoble, France, <sup>9</sup> CeMIA SA, Larissa, Greece, <sup>10</sup> Molecular Genetics Laboratory, Centre for Cardiovascular Surgery and Transplantation Brno and Medical Faculty, Masaryk University, Brno, Czechia, <sup>11</sup> University Clinic of Respiratory and Allergic Diseases Golnik, Golnik, Slovenia, <sup>12</sup> Department of Biophysics, Centre for Research and Genetic Diagnosis of Genetic Diseases, Federal University of São Paulo, São Paulo, Brazil, <sup>13</sup> Department of Immunology & Histocompatibility, School of Health Sciences, Faculty of Medicine, University of Thessaly, Larissa, Greece

## OPEN ACCESS

### Edited by:

Regina Fölster-Holst,  
University Medical Center  
Schleswig-Holstein, Germany

### Reviewed by:

Milos Jesenak,  
Comenius University, Slovakia  
Daniel P. Potaczek,  
University of Marburg, Germany

### \*Correspondence:

Christian Drouet  
christian.drouet@inserm.fr

### Specialty section:

This article was submitted to  
Skin Allergy,  
a section of the journal  
Frontiers in Allergy

**Received:** 14 December 2021

**Accepted:** 16 February 2022

**Published:** 31 March 2022

### Citation:

Drouet C, López-Lera A, Ghannam A, López-Trascasa M, Cichon S, Ponard D, Parsopoulou F, Grombirikova H, Freiburger T, Rijavec M, Veronez CL, Pesquero JB and Germenis AE (2022) SERPING1 Variants and C1-INH Biological Function: A Close Relationship With C1-INH-HAE. *Front. Allergy* 3:835503. doi: 10.3389/falgy.2022.835503

Hereditary angioedema with C1 Inhibitor deficiency (C1-INH-HAE) is caused by a constellation of variants of the *SERPING1* gene ( $n = 809$ ; 1,494 pedigrees), accounting for 86.8% of HAE families, showing a pronounced mutagenic liability of *SERPING1* and pertaining to 5.6% *de novo* variants. C1-INH is the major control serpin of the kallikrein–kinin system (KKS). In addition, C1-INH controls complement C1 and plasminogen activation, both systems contributing to inflammation. Recognizing the failed control of C1s protease or KKS provides the diagnosis of C1-INH-HAE. *SERPING1* variants usually behave in an autosomal-dominant character with an incomplete penetrance and a low prevalence. A great majority of variants (809/893; 90.5%) that were introduced into online database have been considered as pathogenic/likely pathogenic. Haploinsufficiency is a common feature in C1-INH-HAE where a dominant-negative variant product impacts the wild-type allele and renders it inactive. Small (36.2%) and large (8.3%) deletions/duplications are common, with exon 4 as the most affected one. Point substitutions with missense variants (32.2%) are of interest for the serpin structure–function relationship. Canonical splice sites can be affected by variants within introns and exons also (14.3%). For noncanonical sequences, exon skipping has been confirmed by splicing analyses of patients' blood-derived RNAs ( $n = 25$ ). Exonic variants ( $n = 6$ ) can affect exon splicing. Rare deep-intron variants ( $n = 6$ ), putatively acting as pseudo-exon activating mutations, have been characterized as pathogenic. Some variants have been characterized as benign/likely benign/of uncertain significance ( $n = 74$ ). This category includes some homozygous ( $n = 10$ ) or compound heterozygous variants ( $n = 11$ ). They are presenting with minor allele frequency (MAF) below 0.00002 (i.e., lower than C1-INH-HAE frequency), and may be quantitatively unable to cause haploinsufficiency. Rare

benign variants could contribute as disease modifiers. Gonadal mosaicism in C1-INH-HAE is rare and must be distinguished from a *de novo* variant. Situations with paternal or maternal disomy have been recorded ( $n = 3$ ). Genotypes must be interpreted with biological investigation fitting with C1-INH expression and typing. Any *SERPING1* variant reminiscent of the dysfunctional phenotype of serpin with multimerization or latency should be identified as serpinopathy.

**Keywords:** C1-INH-HAE, C1 Inhibitor, *SERPING1* gene, genetic variation, serpin function, serpinopathy, angioedema, hereditary–diagnosis

## INTRODUCTION

Knowledge on angioedema has been first aimed at unraveling the pathophysiology of hereditary angioedema (HAE) with C1 Inhibitor (C1-INH) deficiency, namely, C1-INH-HAE (OMIM #106100; ORPHANET #91,378), a rare genetic disorder that is inherited as an autosomal-dominant trait in most cases. This prototypical condition has been shown to be bradykinin (BK) mediated by a clinical response to a specific BK receptor antagonist (1). Its prevalence is low, from 1/50,000 to 1/100,000, without known ethnic differences (2). HAE pathogenesis has been progressively deciphered from patient observations and basic investigations, showing that HAE results from variants in the *SERPING1* gene in association with the dysfunction of C1-INH (3). When the function of C1-INH has failed, circulating kallikrein–kinin system (KKS) is insufficiently controlled, with subsequent prekallikrein to plasma kallikrein conversion and the production of BK, a vasoactive peptide that causes increased vascular permeability with an activation of the B2 BK receptor. Together with KKS, complement and fibrinolysis are under C1-INH control, all systems sharing many inflammatory features are mediated by the vascular system (4). **Figure 1** shows the central position of C1-INH in the control of kinin forming systems and the additional companions that could be affected by pathogenic variants.

Complement activation is not documented as it is directly involved in HAE pathophysiology. However, plasma kallikrein has been recognized as a pro-convertase, with anaphylatoxin production (6). In addition to anaphylatoxin generation, complement proteases promote plasminogen activation (7), with plasmin production that in turn triggers KKS. This latter proteolytic system directly cleaves circulating high molecular weight kininogen, subsequently generating BK production. Reversely, plasmin has been demonstrated to activate the key complement proteins C3 and C5 (8). These observations provide arguments for an interplay between complement, KKS, and fibrinolysis (4, 9), sustaining inflammation with a pivotal control by C1-INH. Furthermore, medications targeting the KKS or

a B2 BK receptor have been developed and opened a way to understand HAE pathogenesis (10).

An important issue in C1-INH-HAE is the relationship between the systemic plasmatic changes in the KKS activation process and the local effect of BK accumulation in angioedema attacks (11). A failed C1-INH function has been considered as a causative participant. However, other contributors might be critical for the disease severity risk [e.g., kinin catabolism (12) and neutrophil inflammatory mediators (13, 14)] or for an angioedema phenotype of upper airway [e.g., EBV infection (15)]. The contribution of peripheral blood mononuclear cells to the HAE clinical phenotype has been questioned (16). In practice, novel genetic strategies are emerging, resulting in the characterization of a combination of common variants in *SERPING1* and in other genes involved in kinin pathway and metabolism (17).

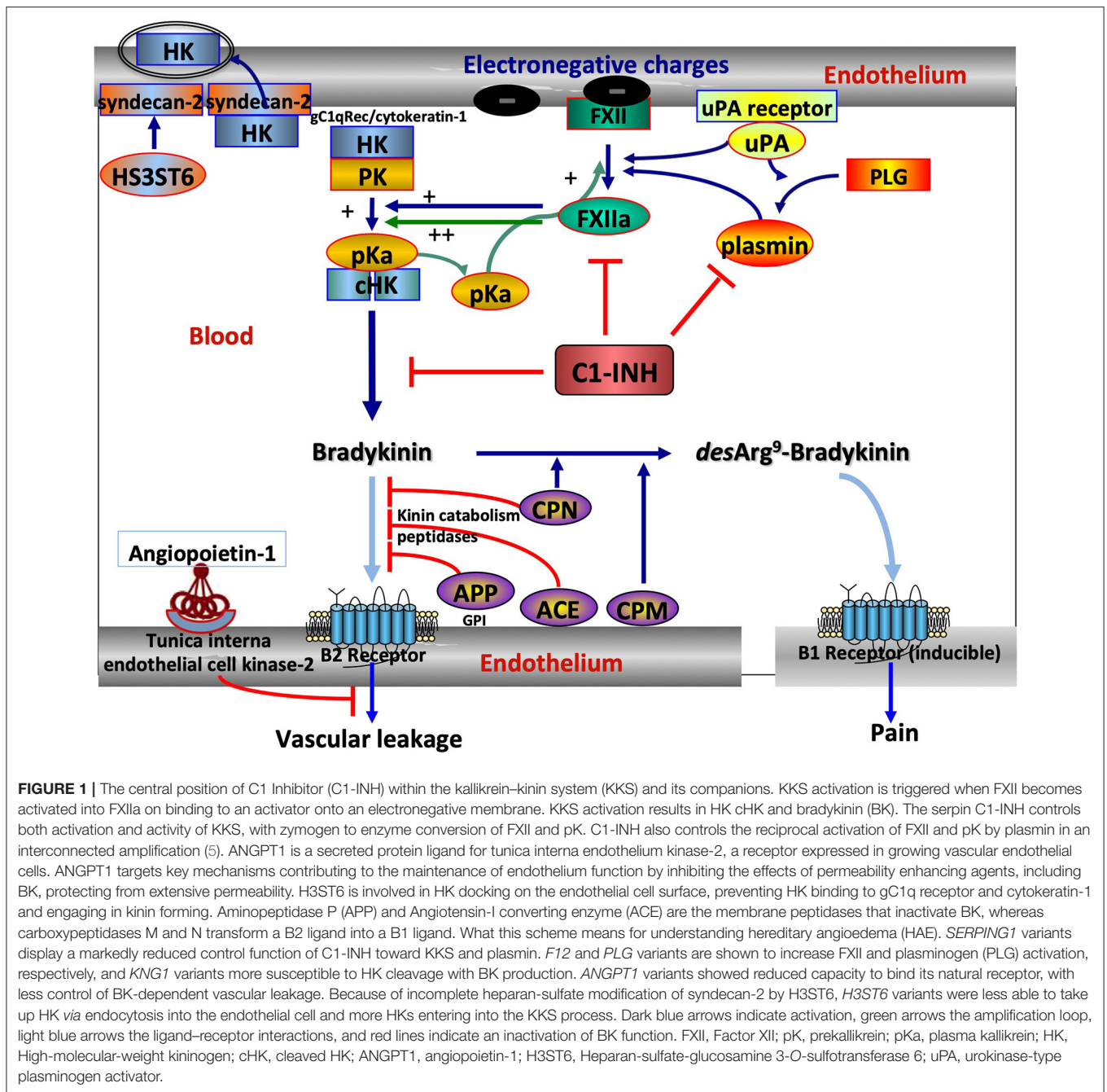
Many observations of families carrying *SERPING1* variants and associated C1-INH data have been collected. This study aims to figure out the state-of-the art of *SERPING1* genetics, with an advantage for C1-INH-HAE diagnostic and its relationship with C1-INH expression and function. Recognizing C1-INH features and *SERPING1* genetics together is a prerequisite for the curation of variant pathogenicity.

## C1-INH DEFICIENCY

The diagnosis of HAE with C1-INH deficiency (C1-INH-HAE) is established on a decreased C1-INH function. Rosen et al. distinguished a type-1 HAE (HAE-1), where C1-INH-HAE results from the failure to synthesize the protein, from a type-2 HAE (HAE-2) where an abnormal, dysfunctional protein is synthesized (18). HAE-2 is commonly identified from the data presenting with normal, or elevated, antigenic C1-INH in serum (19). However, many dysfunctional missense variants with a low antigenic C1-INH have been shown to be expressed together with the normal allele (20); they are characterized as HAE-2. Sharing same clinical presentation, diagnostic and pathophysiology, both types are also sharing the same recommendations for treatment options (10). This suggests that a distinction between HAE-1 and HAE-2 should not be relevant for physician tasks, but could be valuable for a structure–function relationship in the identification of pedigrees.

Substantial misdiagnoses and the delayed diagnoses of C1-INH-HAE are common. A significant obstacle for diagnosis is its low prevalence, high phenotypic variability, and incomplete

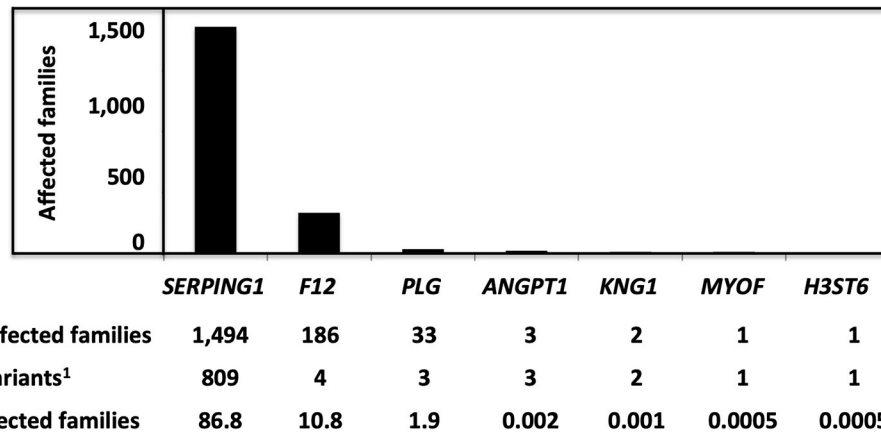
**Abbreviations:** ACMG, American College of Medical Genetics; BK, bradykinin; C1-INH, C1 Inhibitor; C1-INH-HAE, Hereditary angioedema due to C1 Inhibitor deficiency; HAE, Hereditary angioedema; HAE-1, Hereditary angioedema type 1; HAE-2, Hereditary angioedema type 2; KKS, Kallikrein–kinin system; MAF, Minor Allele Frequency; nC1-INH-HAE, HAE with normal C1-INH function; RCL, Reactive center loop; SRE, Splicing regulatory element; VUS, variant of uncertain significance.



penetrance, with the failed suspicion of angioedema symptoms, in particular abdominal attacks (21). This emphasizes the need to identify patients with HAE and HAE families to improve disease management and patient outcome (22).

Hereditary angioedema due to C1 Inhibitor deficiency shares a common kinin dependency with other HAE situations: F12-HAE, PLG-HAE, KNG1-HAE, ANGPT1-HAE, and HS3ST6-HAE, but not with MYOF-HAE, U-HAE (Figure 1). The distribution of HAE conditions is shown in Figure 2. With 1,494 affected families recorded so far,

C1-INH-HAE represents a predominant HAE condition (86.8%). Variants of these HAE-associated genes have been characterized with autosomal-dominant transmission and incomplete penetrance. Some rare *SERPING1* variants have been found in affected families with a recessive transmission (Section Recessive Variants), and *de novo* variants have been identified in 5.6% of the probands (20). These C1-INH-HAE features prompt biologists to perform the most accurate assays of the function of C1-INH for analytical HAE diagnostics.



**FIGURE 2 |** The distribution of variants responsible for HAE and characterized as pathogenic/likely pathogenic, with a number of affected families. Pathogenic/likely pathogenic variants have been characterized in agreement with the American College of Medical Genetics (ACMG) criteria or as declared by authors in the observations. <sup>1</sup>Variants were characterized as pathogenic/likely pathogenic.

## The Function of C1-INH and Its Analysis in Plasma

C1 Inhibitor is a multifunctional serine protease inhibitor (serpin) that controls various serine proteases involved in multiple plasmatic cascades [e.g., KKS, thereby limiting the production of the vasoactive peptide BK (23)]. C1-INH is a single-chain, highly glycosylated circulating protein of Mr 105 kDa (nonreducing conditions) and 478 aminoacid residues (24).

As a serpin (clade G), it regulates serine proteases *via* an irreversible suicide-substrate mechanism as a set mousetrap (25). C1-INH consists of 3  $\beta$ -sheet structures, 9  $\alpha$ -helices, and a Reactive Center Loop (RCL) located at the top of the central  $\beta$ -sheet (**Figure 3A**). RCL serves as a bait region for specific proteases. The target protease recognizes and cleaves the P1-P1' scissile bond in the RCL, with an insertion of the hinge and RCL into the central  $\beta$ -sheet A as an additional strand 4A. This drives the covalently bound protease to the base of the serpin molecule (26, 27). After this conformational change, the serpin adopts a thermodynamically stable conformation, with an irreversible inhibition of the protease. C1-INH-protease complexes are then cleared by the liver (28). This remarkable conformational change represents a common feature for serpins and illustrates a conformational pathology (25, 29). C1-INH also directly interacts with native C1 to prevent the autoactivation of C1 (30), thereby inhibiting its own consumption.

Laboratory identification of C1-INH-HAE is challenging with a primary interest for physicians, with the need (i) to recognize a C1-INH dysfunction and (ii) to decide on a molecular diagnosis, e.g., for patients with HAE without family history or with records of inconsistent biochemical measurements.

C1-INH Inhibitor circulates in the plasma (0.21–0.35 g/L), corresponding to a nearly 100% serpin function measurement. Two types of C1-INH function assays are available, the residual enzymatic C1s protease/KKS activity (chromogenic assay) or the C1-INH-protease complex formation using an ELISA. The

chromogenic assay is more sensitive than the ELISA for detecting a low C1-INH function and is often preferred (31, 32).

The genetic basis of C1-INH-HAE means that family history represents a starting point for diagnosis. So, all family members must be strongly encouraged to be tested when a C1-INH-HAE has been diagnosed in a relative.

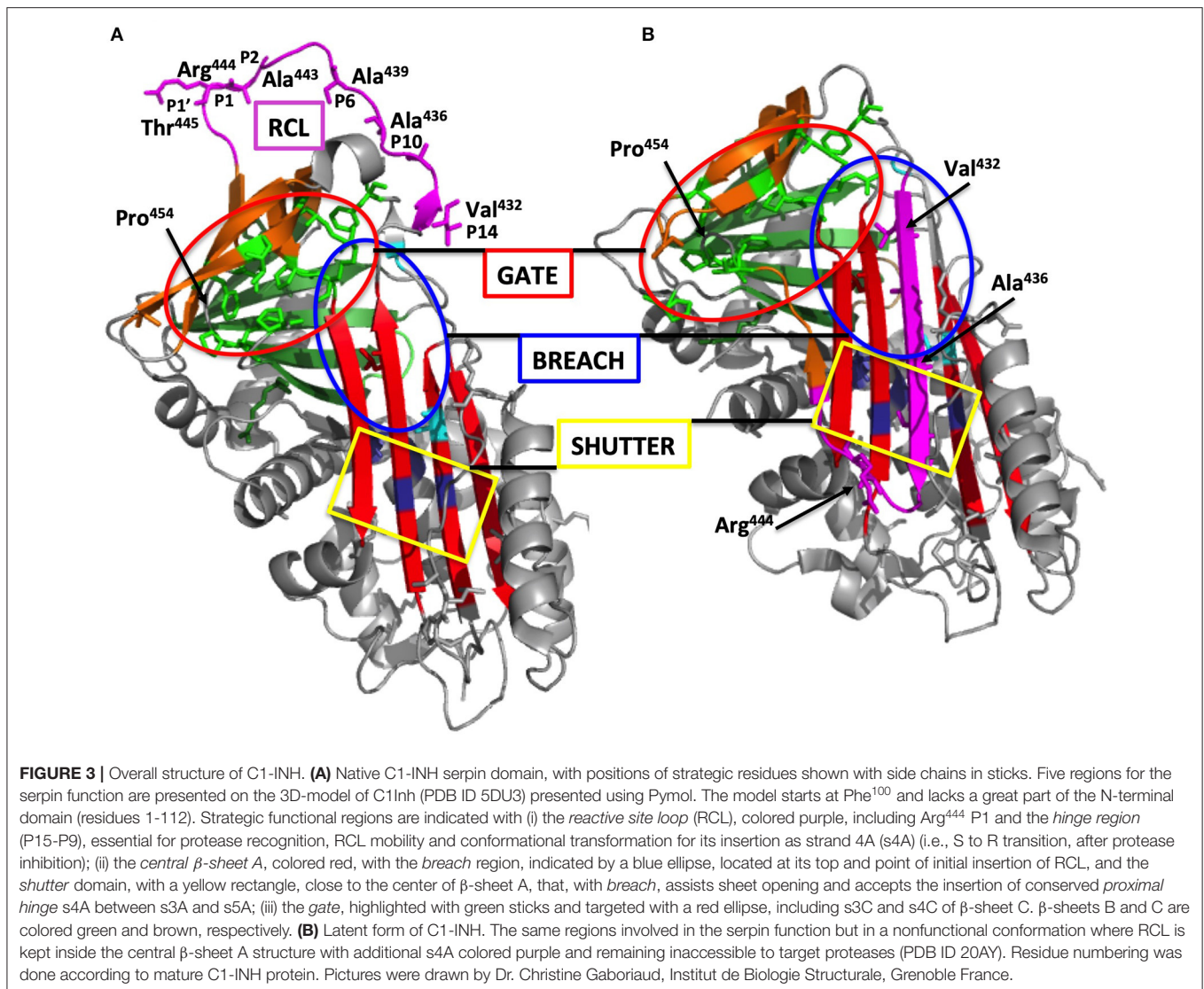
## C1-INH Expression Criteria: Biological Phenotype

Variants commonly characterized as HAE-2 from the only normal antigenic C1-INH criterion are found within the RCL (i.e., variants affecting the positions Ala<sup>443</sup>-Arg<sup>444</sup> in the mature sequence; **Figure 3A**) and at the positions Gln<sup>201</sup> and Lys<sup>251</sup> (20). Many missense variations meet the Rosen's criterion of HAE-2 (18) and affect the functionality after protein misfolding. When an abnormal, nonfunctional protein is synthesized, circulating C1-INH presents with a remnant 105-kDa species after incubation with equimolar C1s protease on the anti-C1-INH immunoblot (20). This analytical strategy is helpful to identify the molecular features of any missense variant, with HAE-2 characteristics, including latent and oligomerized forms (33).

## Haploinsufficiency

As C1-INH-HAE is inherited as a dominant disorder in heterozygous cases, with one normal allele, antigenic C1-INH should theoretically be 50%. However, common observations recognized that C1-INH values are <35% of normal (i.e., from analytical threshold to 35%); attacks of angioedema are likely to occur when functional C1-INH levels are within this range (34).

Haploinsufficiency is common in C1-INH-HAE, where a dominant-negative variant product impairs the expression of normal allele. Haploinsufficiency may occur either through a *trans*-inhibition of normal protein expression (e.g., in an intracellular retention subsequent to intermolecular aggregates),



or with decreased C1-INH production due to altered epigenetic control (Section Dominant-Negative Effect).

Decreased C1-INH function in plasma could be caused by high catabolism of remaining C1-INH was demonstrated in a 95-kDa species in patient plasma (20). This molecular distribution is in line with an increased proteolysis, rather than with a haploinsufficiency.

### Limits of Biological Testing

Transient decreases of C1-INH function are recurrently found in *F12*-c.983C>A;p.(Thr328Lys) female carriers, with angioedema attacks precipitated by estrogen intake or pregnancy. This decrease is consistent with C1-INH proteolysis by activated KKS (35).

Variant identification could be essential for the HAE diagnosis of patients presenting with symptoms, but also with the biological features of AAE-C1-INH or autoimmunity and with the lack of HAE family history, as reported by Veronez (36).

### Homozygous and Compound Heterozygous Proband Carrying SERPING1 Variants

Hereditary angioedema due to C1 Inhibitor deficiency dominant trait, though with incomplete penetrance in heterozygous probands, early suggested that homozygosity for *SERPING1* variants may be embryonically lethal despite the fact that no signs of increased developmental lethality are present in HAE cohorts (37). In the last 15 years, several HAE pedigrees evidencing a recessive manner of inheritance have been documented and to date account for 10 true homozygous probands and 11 compound heterozygote alleles segregating in nine pedigrees. In general, *SERPING1* variants with recessive behavior represent a minor percentage of the total (5.8%) and are commonly classified by pathogenicity prediction tools such as “benign,” “likely benign,” or “variant of uncertain significance (VUS).” They are rare in the general population, usually presenting with minor allele frequencies (MAFs) below 0.0001. No *SERPING1*

region has been recognized as specifically linked to a recessive feature, the variants being distributed all over the gene. No specific mutation categories are statistically associated with homozygosity, with two promoter, six missense, one splicing, and one indel variants (**Supplementary Table S1**).

The variants c.[-(163)C>T] and c.[-(161)A>G] are located in the promoter region of *SERPING1* and disrupt a putative CAAT box located at -62 bp from the origin of transcription (38–40). These two variants provide a hypothetical mechanism for their pathogenicity based on the defective transcription of *SERPING1*. The disruption of CAAT promoter sequences hampers the transcription of the affected alleles due to a defective binding of the RNA polymerase II and other nuclear-binding factors. However, in the case of *SERPING1*, there exists no direct evidence of the functionality of its CAAT sequence and therefore additional mechanisms may be postulated (41). The remaining recessive *SERPING1* variants reported affect the coding sequence of the gene and account for one variant in the 5' untranslated region and six variants distributed through exons 4, 7, and 8. The variant c.-21T>C in the second nucleotide of exon 2 was reported as a probable cause of the disease in a homozygous proband with a HAE-1 phenotype (42). Due to its relatively high MAF (0.03), it has been reported as a polymorphic or disease-modifying allele (38, 43–45).

The seven remaining homozygous variants located in the coding sequence are associated with a HAE-1 phenotype in symptomatic homozygotes and with a HAE-2 (most commonly) or an asymptomatic presentation in heterozygous carriers. An illustrative example of this segregation of phenotypes is the c.1385T>G;p.(Ile462Ser) variant. In the original pedigree described by Blanch et al., HAE manifested exclusively in one of the two homozygous siblings as a HAE-1 phenotype while all their heterozygous relatives remained asymptomatic despite presenting with C4 consumption and a low C1-INH function (37). Interestingly, the homozygous probands from this family, as well as those with the c.[1198C>T];p.(Arg400Cys) (46) and c.[1379C>T];p.(Ser460Phe) (47) variants, presented with a very low or an undetectable antigenic C1q, thus reminding the phenotype of some patients with acquired angioedema with C1-INH deficiency. This acquired C1q deficiency is not an invariant trait in patients with C1-INH homozygous deficiency but pertains to the specific functional impairments of some recessive variants (47). Other variants have been found in homozygous carriers (**Supplementary Table S1**): c.[440T>A];p.(Val147Gln), c.[668A>C];p.(Gln223Pro), c.1202T>C;p.(Ile401Thr), and [c.646\_647delAinsTCAGTGTCTGTG], and the latter is characterized as a *de novo* variant (48).

Compound heterozygosity of *SERPING1* alleles with highly variable and incomplete penetrance is also a cause of C1-INH-HAE, with pedigrees presenting with uncommon symptomatic individuals (**Supplementary Table S1**); one or both allele(s) has (have) been characterized as VUS/benign variant(s).

## Digenic Variants

On the other hand, the combined presence of a *SERPING1* variant and a variant in another HAE susceptibility gene is now recognized as a cause of the disease. Examples of such

situations are the combination of *SERPING1* variants with the *F12*-c.-4C>T polymorphic variant (rs1801020; MAF 0.472) (49, 50) and that of the c.513 + A>G *SERPING1* variant with the *F12*-c.1032C>A;p.(Thr328Lys) pathogenic variant presenting HAE symptoms since 7 years of age with a severe phenotype (51).

A curious case shows an interesting pattern of the localization of swellings in a family composed by patients carrying (i) the variant c.1480C>T;p.(Arg494\*) in *SERPING1* in heterozygosis, (ii) the variant c.988A>G;p.(Thr328Lys) in *PLG* in heterozygosis, or (iii) both (52). Individuals carrying only the *SERPING1* mutation or the combination of *SERPING1* and *PLG* mutations presented abdominal pain and edema of the extremities (hands and feet) more frequently when compared to the patients carrying only the *PLG* variant. Records of patients with *PLG*-HAE present with a higher proportion of attacks affecting the tongue and face, and less abdominal attacks (53), which was confirmed in this family case study (52). The presence of *SERPING1* and *PLG* pathogenic variants demonstrated a combination of symptoms but was not enough to prove an increase in the severity of the disease phenotype.

In an attempt to explain and correlate the variability in the manifestation of symptoms in C1-INH-HAE with other genes, Veronez et al. (54) evaluated 45 *SERPING1* mutation carriers and 15 healthy relatives from 26 families. The authors analyzed 15 genes entangled in the function of KKS and metabolism of associated enzymes and ligands/receptors using a next-generation sequencing (NGS) panel, and a total of 211 different variants were identified in the 15 genes analyzed. *BDKRB2* and *CPM* presented a large number of variants in untranslated regions, whereas *ACE*, *CPM*, and *NOS3* genes presented a higher number of variants directly affecting amino acid sequence. Despite the large amount of variants identified, no specific variant was significantly associated to any of the clinical symptoms affecting the patients (facial, abdominal, extremities, upper airways, and genitalia), indicating that the modulation of HAE symptoms could require a more complex regulation, probably involving pathways beyond the KKS, epigenetics, and environmental factors.

Aiming to uncover the genetic basis of nl-C1-INH-HAE, Loules et al. also applied a custom NGS platform to analyze 55 genes related to KKS involved in angioedema pathogenesis (55). Patients with normal antigenic C1-INH were evaluated in the study using patients with C1-INH-HAE as control, deciphering the presence of common variants that could modulate the patient clinical phenotype. Although the frequency of variants per gene was comparable between HAE with normal C1-INH function (nC1-INH-HAE) and C1-INH-HAE, variants of the *KNG1* and *XPNPEP1* genes were detected only in patients with nC1-INH-HAE. The authors concluded that alterations in some genes (e.g., *KNG1*) could play a role in the complex trait of HAE. These results emphasize the importance of modulator genes in HAE clinical expression with a better understanding of disease pathophysiology. This observation possibly drives the discovery of new therapeutic targets and provides useful indicators for disease clinical management.

## Polygenic/Oligogenic Conditions

Recently, the results suggest that polygenic situation is common in C1-INH-HAE and can significantly influence the penetrance of the disease, as exemplified by Veronez et al. (56). Almost all symptomatic patients from a pedigree presenting with C1-INH deficiency due to the c.1369G>C;p.(Ala457Pro) *SERPING1* variant carry multiple allele combinations with *ACE* [c.970C>T;p.(Arg324Trp)], *ENPEP* [c.638A>G;p.(Gln213Arg)], *KLK1* [c.433G>C;p.(Glu145Gln) or c.556A>G;p.(Lys186Glu)], *KLKB1* (c.428G>A;p.(Ser143Asn) or c.1679G>A;p.(Arg560Gln)], *KNG1* (c.533T>C;p.(Met178Thr) or c.591T>G;p.(Ile197Met)], *NOS3* (c.894T>G;p.(Glu298Asp) or c.2654G>T;p.(Arg885Met)] or *PRCP* (c.336A>T;p.(Glu112Asp)] alleles (56). This genetic complexity is pertaining with the present understanding of HAE as a pathology caused by an overactivation of the KKS and/or by a kinin accumulation.

## DISTRIBUTION OF VARIANTS

The *ensembl* database displays 8,574 variants for the *SERPING1* gene (URL [www.ensembl.org](http://www.ensembl.org); ID ENSG00000149131; retrieved on October 28, 2021), with a homogeneous distribution along the 21,785 bases (GRCh38/hg38; latest assembly). Only 809 have been registered as pathogenic or likely pathogenic and 50 as VUS, according to the published observations and fulfilling the American College of Medical Genetics (ACMG) recommendations (57, 58). These variations are recorded in genetic databases (e.g., LOVD, tracked October 28, 2021; ClinVar, tracked November 2, 2021). **Supplementary Table S2** shows the distribution of pathogenic/likely pathogenic variants.

## Missense Variants

Taking into account pathogenic/likely pathogenic/VUS variants, the distribution of missense variants is highly unbalanced between the mucin-like N-terminal domain (8/112 aminoacid residues; i.e., 7.1% of residues are affected) and the serpin domain (241/366; 65.8%); six variants have been recognized within the signal peptide sequence (6/22; 27.2%). This is congruent with a great impact of any modification of the peptide sequence within the serpin domain on C1-INH dysfunction. This prompts the data curator to a special attention for the interpretation of a missense variant with functional structures of strategic importance for serpin biology: gate, shutter, breach, hinge region, and less importantly, polyanion-binding domain (**Figure 3A**).

Missense variants (32.2% of all disease-causing variants) must be considered in association with mutant allele expression and C1-INH function. Most of the variant products are expressed, fulfilling criteria of a HAE-2 or an intermediate type, and have been distinguished between class I (i.e., altered exposure of the active site), class II (i.e., disturbed insertion of the RCL), and class III (i.e., conformational transition with spontaneous self or mutual insertion of the RCL) (20, 59) (**Figures 2, 3**); the latter could shape a M\* serpin conformation prior to the formation of extending chains of ordered and thermodynamically stable polymers, a characteristic of serpinopathy, or C1-INH species taking a stable and pathological latent form (**Figure 3B**) (25).

## Large Deletions/Insertions

Large deletions/duplications are likely to result from the recombination between *Alu* repeat sequences present in most introns of the *SERPING1* gene (60), and account for 8.3% of all disease-causing variants in the *SERPING1* gene (19; LOVD). The 7 introns contain 19 *Alu* repeat sequences, and a high density of *Alu* repeats has been reported in introns 3, 4, and 6. This is considered as a hotspot for nonhomologous recombination resulting in deletions or duplications, with exon 4 as the most affected (22, 60). Different approaches, including Southern blot analysis (61), fluorescent multiplex PCR (62–64), and long-range PCR (22, 65), have been used to detect those variants, while MLPA represents the reference technique for the detection of large deletions and insertions (22, 64, 65). The disadvantage of those methods is that it cannot precisely locate the boundaries of the insertion/deletion. To determine a precise location, time-demanding combination of previously mentioned methods with direct sequencing of the boundaries must be performed (64–66). Recently, a targeted NGS platform has been developed and validated to simultaneously detect the large deletions/insertions providing at the same time both exact size and location of the deletions/insertions (67). Up to now 64 large deletions/insertions in the *SERPING1* gene have been reported (19; LOVD), but for the majority, the exact size and location of boundaries remain unknown.

## Small Deletions/Insertions

Small deletions/duplications/insertions are abundant in *SERPING1* variants (36.2%) and mostly with subsequent frameshift and characterized as pathogenic/likely pathogenic.

## Variants Affecting Splicing

Splicing affecting variants typically have a deleterious effect on protein expression and/or function. They can result in exon skipping (or even multiple exon skipping), intron retention, *de novo* splice site creation, cryptic splice site usage, or a combination of 2 or more of these effects. All mentioned options were demonstrated to occur in *SERPING1* variants (68).

## Variants at Intron/Exon Boundaries

Splicing mutations comprise 14.3% of *SERPING1* mutations, and 47% of them are located in conserved canonical positions ( $\pm 1$ ,  $\pm 2$ ) of splice sites adjoining all exons of the gene (20) (**Supplementary Table S5**). According to HGMD and LOVD databases, 35 substitutions and 13 deletions and insertions changing canonical positions have been described as HAE-1-causing variants. Pathogenicity of these variants is well established and easily assessed by *in silico* prediction tools.

The evaluation of potentially pathogenic splicing variants in other than canonical positions is more difficult and considerably less reliable using *in silico* prediction tools; especially of those affecting splicing regulatory elements (SRE). Importantly, various procedures have been applied to prove their effect on splicing, from analyses of *SERPING1* variant transcript from patients' blood-derived RNA samples or from minigene splicing assays (62, 67).

Siddique et al. referred for the first time the correlation of an intronic variant with HAE in a family carrying c.10291 + G>T (69). The function of the variant could not be proven as the mRNA appeared to be normal. However, the wild-type mRNA had a relative abundance of  $\approx 50\%$ , suggesting that the mutant replicate was not converted to a stable mRNA and rapidly degraded.

Up to now, 31 potentially pathogenic variants in noncanonical intronic positions of splice sites have been described. Whereas pathological impact of some of these variants has been well established based on functional tests or affirmative results of multiple prediction tools and/or the presence in several patients, other variants require further analysis of their functional significance. Illustrative examples are given by c.686-12A>G (44, 70), creating a *de novo* splice site in intron 4 (68) (**Figure 4**), c.52-10T>A in intron 2 after the observation of alternative short transcript species (72), and c.1250-13G>A in intron 7 after an *in silico* analysis (40).

Consequences of several new splicing variants already reported (20) could be thus evaluated as pathogenic/likely pathogenic, and the effect of others (e.g., c.890-8C>G) remains to be further established.

Interestingly, Colobran et al. detected c.6852 + T>A in intron 4. Functional studies of the mRNA demonstrated that this variant leads to the omission of exon 4 (73). Exon 4 consists of 135 bp (i.e., 45 codons); the lack of exon 4 corresponds to an in-frame deletion. Thus, the mutant allele could produce a protein lacking 45 amino acids. The levels of the mutant mRNA have been found to be very low compared to the wild-type, indicating that the mutant mRNA was degraded by one of the three surveillance pathways. In addition, the bioinformatic tool RNAfold® predicted a modification of the secondary structure. These data are consistent with the degradation of the mutant mRNA *via* the no-go degradation (NGD) pathway, which is associated with secondary structural features.

In  $\approx 85\%$  of cases, a G base is located in the fifth nucleotide of an intron (74). Variants at the +5 position are thought to significantly reduce the binding at the 5' splice site to the complementary site at the U1snRNP particle, one of the first steps in the complex process of mRNA splicing (75). Consequently, the immediately preceding exon is omitted, followed by the activation of a deviating 5' splice site and complete retention of introns (76). Alternatively, variants at this site may result in a reduced quantity of wild-type mRNA, or qualitative defects, by omitting an exon, activating a cryptic splice site, or creating a new splice site. The variant c.-22-19\_-22-4del was detected in intron 1, predicted as pathogenic by the bioinformatic tool MutationTaster®, but marked as VUS, until a functional study and/or other evidence proves its pathogenic significance (77). At c.550-5 in intron 3, both variants c.551-5T>A and c.550T>G have been characterized as pathogenic due to its location near the 3' splice site (**Supplementary Table S5**). The c.890-14C>G variant has been detected in intron 5 (78) and the c.1250-13G>A variant in intron 7 (40, 79), the latter reducing the possibility of splicing at a rate similar to a variant located at the canonical positions. Other examples are displayed in

**Supplementary Table S5:** c.515 + G>A, c.5505 + G>C, c.5505 + G>A, c.12495 + G>A, and c.12495 + G>T.

Exonic variants can also affect splicing. The impact of several exonic variants on splicing has been functionally evaluated using the RNA analysis. Three substitutions in the last nucleotide of the exon 3 (position c.550), the deletion of the last nucleotide of the exon 4 (c.685del) (**Figure 5**), and substitutions in the proximity of the 3' end of the exon 5 (c.882C>G, c.884T>G) have been shown pathogenic (63, 68). Moreover, the exonic variant c.-21T>C activates a cryptic acceptor site and causes exon 2 skipping to a certain extent (80) and might be linked to a more severe phenotype when occurring in *trans* with another pathogenic variant (20, 80).

### Deep-Intronic Variants (> 20 nt Upstream/Downstream of the Exon)

Near the 3' splice site, a pattern of polypyrimidines (C or T, 5–40 nucleotides) is usually located between the branch point and the 3' splice site. As stated in Section Variants at Intron/Exon Boundaries, consequences of new variants within introns need additional consideration, in particular, the influence of the c.6853 + 1G>A variant (72).

Four variants have been detected deeper within the *SERPING1* gene and have been associated with HAE, two being considered as deep-intronic variants (i.e., located  $\approx 100$  bp from the intron–exon junction). The variant c.1029 + 84G>A in intron 6 is recurrent, detected in the affected families, and thought to affect the transcriptional process (44, 81–84). The bioinformatics analysis predicted that c.1029 + 384A>G results in a *neo*-5' splice site with the incorporation of a pseudo-exon into the transcript. The inclusion of the pseudo-exon leads to an early termination codon and possibly to the destruction of the produced mRNA by cell control mechanisms, which could potentially explain the reduced amount of proteins. The functional study performed by RT-PCR finally proved the above prediction (83). Vatsiou et al. detected the variant c.-22-155G>T in intron 1 in four patients (85). The *in silico* analysis predicted that the variant c.-22-155G>T causes a deleterious effect on the gene and degradation of the mutated transcript by the mRNA surveillance pathways; barring this variant was demonstrated by the loss of heterozygosity on the cDNA level. The variant was classified as pathogenic, in line with ACMG-AMP 2015 guidelines (e.g., Intervar) (57).

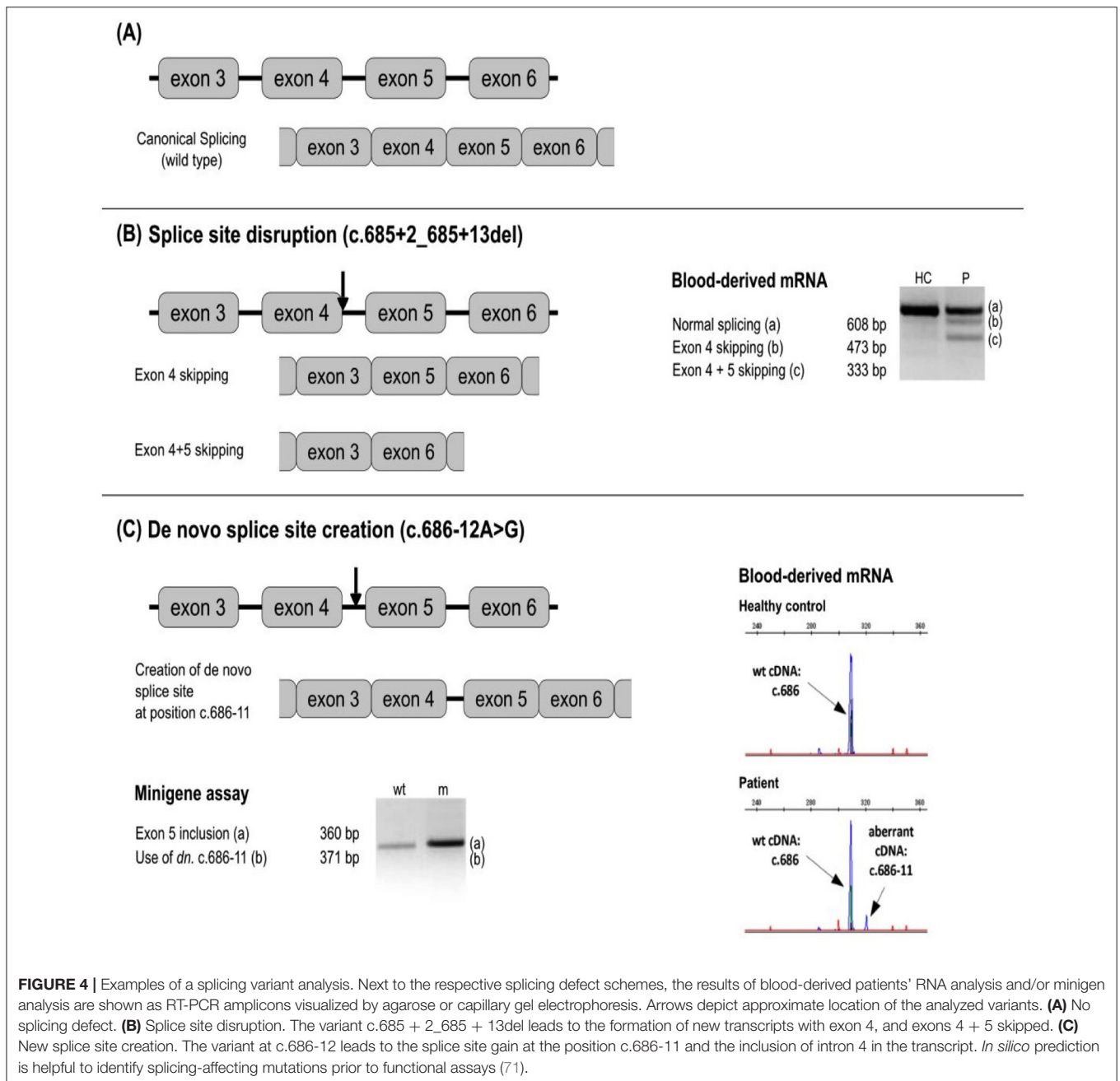
## INHERITANCE OF VARIANTS

### Dominant-Negative Effect

Dominant-negative involves a change of the function; the disease is not caused due to the loss of protein function, but happens due to a change in protein function or in disequibrated mRNA species between alleles. The dominant-negative variant acts antagonistically in the wild-type allele by impairing its expression or by biochemically interacting with the normal gene product and interfering with C1-INH function.

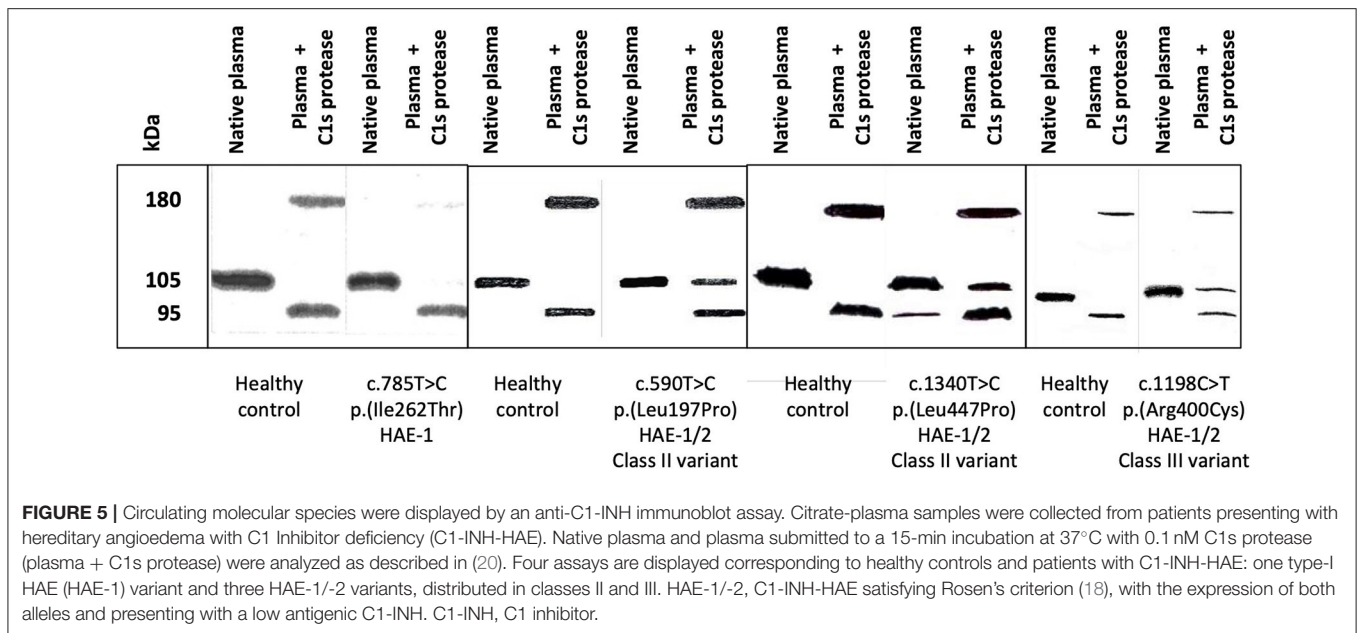
A first pioneering investigation on C1-INH transcripts has shown a “*trans*” inhibition of the normal allele by variant mRNA or protein (86). The authors studied C1-INH expression in





fibroblasts in which the mutant and wild-type mRNA and protein could be distinguished because of the deletion of exon 7. In patient cells, the wild-type mRNA was expressed at  $\approx 50\%$  of normal, whereas the mutant mRNA was 17% of normal. Rates of synthesis of both normal and mutant proteins were 11% and 3% of normal, respectively (i.e., lower than predicted from the mRNA levels), suggesting a *trans*-inhibition of normal allele by a variant product. A substantial reduction of both mRNA as well as C1-INH protein expression has also been shown using two allele-specific PCR in a carrier of the variant c.\*101\_\*254del, a 155-bp deletion 100 bp downstream of the stop codon in exon 8 (87).

An elegant demonstration of dominant-negative effect of variants has been provided by Haslund et al. (88), where the mutated C1-INH species affected wild-type C1-INH in a dominant-negative manner with intracellular/plasmatic C1-INH aggregates and subsequent reduction in functional C1-INH. C1-INH encoded by a subset of HAE-causing *SERPING1* alleles disturbed the secretion of normal C1-INH protein in a dominant-negative fashion by triggering the formation of protein-protein interactions between normal and mutant C1-INH. The authors observed large intracellular C1-INH aggregates that were trapped in the endoplasmic reticulum.



Interestingly, the transfection of wild-type *SERPING1* constructs into cells was able to remove the block in normal C1-INH secretion (88). Polymerogenic variants have been recognized with a location in proximity to/within the insertion site of RCL in C1-INH (i.e., shutter and hinge/gate regions; **Supplementary Table S3**), for example within  $\alpha$ -helix C, c.566C>A;p.(Thr189Asn),  $\beta$ -sheet 3A, c.878T>C;p.(Ile293Thr) (33),  $\alpha$ -helix F, c.838\_846del;p.(Leu281\_Ser283del) (88). The first variant prone to oligomerization has been recognized with c.1372G>A;p.(Ala458Thr) within RCL (89) and c.818\_820del;p.(Lys273del) affecting the inter-domain  $\alpha$ -helix F/ $\beta$ -sheet 3A (90). More recently, an additional variant p.(Ser150Phe) targeting  $\alpha$ -helix A has been shown to be stably expressed within the cultured cells and not secreted into the medium at all. The mutant C1-INH significantly prevented the secretion of wild-type C1-INH, with its degradation within the cytoplasm through an interaction with the mutant protein (91). Observations of protease-resistant mutant C1-INH species suggest circulating latent C1-INH species that could also represent stable and low energetic conformations (**Figure 3B**, **Supplementary Table S3**) (20, 92).

## Recessive Variants

Some variants have been shown to circulate in a 50% expression of normal C1-INH allele, with an absence of haploinsufficiency (**Supplementary Table S4**). They do not segregate with symptomatic carriers. Homozygous carriers have been recognized as symptomatic for angioedema or these variants may have a disease-modifying effect (Section Homozygous and Compound Heterozygous Proband Carrying *SERPING1* Variants).

The recurrent c.1198C>T;p.(Arg400Cys) variant has been studied for its remittent expression (93). It is responsible for a temporary drop of C1-INH function with an enhanced effect

in a homozygous state to express a HAE phenotype (recessive inheritance; 45). Once correctly folded, C1-INH<sup>Arg378Cys</sup> is secreted as an active, although quite unstable, monomer. However, it could bear a folding defect, occasionally promoting protein oligomerization and interfering with the secretion process. Environmental factors (i.e., temperature, pH, and oxidative stress), which could apply even in situations of mild physical stress, like hyperthermia or metabolic acidosis, have been demonstrated *in vitro* to affect the stability and the function of C1-INH<sup>Arg378Cys</sup>.

Rare variants of *SERPING1* yielding deficient inhibitory activity toward complement C1 proteases but not toward the KKS proteases result in paucisymptomatic HAE. These have been characterized within the RCL [e.g., variations at the P2 position c.1394C>T;p.(Ala465Val) (94), at the P6 position c.1382C>T;p.(Ala461Val) (95, 96)], or out of the RCL [e.g., c.452T>G;p.(Leu151Arg) (20, 97)]. The position P2 in C1-INH supports protease specificity, with a significant reduction in rate constants for the reaction of C1-INH<sup>Ala443Val</sup> with C1r, but not with C1s, FXIIa, plasma kallikrein or plasmin (96). However, C1-INH<sup>Ala439Val</sup> (i.e., a variation at P6) displays a moderate decrease in control of C1s, compared with the wild-type protein (96).

## Uniparental Disomy

Gonadal mosaicism situations have rarely been detected in C1-INH-HAE; this condition must be distinguished from a *de novo* variant. The identification of mosaicism is important in establishing the disease diagnosis, assessing recurrence risk and genetic counseling. The pathogenic variant could be inherited from a maternal or paternal allele.

## From a Maternal Allele

A c.[597C=>G] condition has been found in a family in which only both sons, but not the parents, show clinical and laboratory

findings typical of HAE, with allele segregation demonstrated using DHPLC (98). c.597C>G variant has not been detected in DNA derived from buccal cells, urinary cells, hair roots, and cultured fibroblasts from the mother, whereas it has occurred on the maternal transmitted chromosome.

### From a Paternal Allele

- (1) Family 1. A c.[69\_139=]/del has been shown in a family where three sons, but not the parents, show clinical and laboratory findings typical of HAE, with demonstrated allele segregation (99). c.69\_139del variant has not been detected in DNA derived from somatic cells from the father and the mother, whereas it has occurred on the paternal transmitted chromosome.
- (2) Family 2. A c.[536C=>T] has been observed in a family in which only both sisters, but not the parents, show clinical and laboratory findings typical of HAE, with allele segregation demonstrated using Sanger sequencing (100). c.536C>T variant has not been detected in DNA derived from lymphocytes from the father and the mother, whereas it is present on the DNA prepared from the sperm of the father and on the paternal transmitted chromosome.

## NONCAUSAL VARIANTS AS GENE MODIFIERS

The combination of genetic variants may explain the variability in the manifestation of symptoms in C1-INH-HAE, in which nonpathogenic variants in diverse genes may confer susceptibility to a more severe phenotype when associated to pathogenic mutations in *SERPING1*. Within *SERPING1*, the c.-21T>C variant has been recognized as a disease modifier (Sections Homozygous and Compound Heterozygous Proband Carrying *SERPING1* Variants and Variants at Intron/Exon Boundaries). Beyond the known pathogenic mutations in *F12* gene that are described as causative for HAE (c.983C>A, c.983C>G, c.971\_10182 + 4del, c.892\_909dup), a common polymorphism, c.-4T>C (*F12*-46C/T) is demonstrated to influence the severity of disease in patients with C1-INH-HAE (50). This variant was associated with a delay in the onset of symptoms and with a decreased necessity to use long-term prophylaxis therapy (50, 101). A further study confirmed low FXII serum levels in patients with C1-INH-HAE carrying the T allele and found that asymptomatic patients presented the T allele in a higher frequency compared to symptomatic ones (102). Another study sequenced the exonic and regulatory regions (5'-UTR and 3'-UTR) of *F12* gene from 161 C1-INH-HAE and 191 HAE-nC1-INH, and found 6 *F12* polymorphisms in patients with C1-INH-HAE and 9 in patients with HAE-nC1-INH, including rare and first described variants (103). Variants such as *F12*-c.1768T>G;p.(Cys590Gly), which affect the catalytic domain of *F12* in a hotspot previously associated with protein deficiency, could be beneficial to HAE genetics. However, more studies are needed to establish any protective association of this variant in HAE (54).

## DATA CURATION

### *In silico* Rating

Although the genetic screening of the *SERPING1* gene has been facilitated by recent high-throughput technologies that have been available for massive DNA sequencing, clinical classification of the detected variants remains challenging. *SERPING1* variants could be divided into categories according to their possible pathogenicity:

- All variants (nonsense, frameshift, splicing, and large defects) with structural changes or misfolding of the protein, likely associated with a deleterious impact.
- Missense variants and changes affecting untranslated sequences in the 5' or 3' ends, both lacking strong evidence regarding their pathogenicity.

Pathogenicity supporting evidence provided by large pedigrees is required, such as functional analyses, population data, *in silico* predictions, and segregation family studies. Bioinformatic tools (e.g., SIFT®, PolyPhen®, Mutation Taster®) have been developed based on evolutionary conservation, the type of amino acid change, and the position within a functional domain, allele frequency. The certainty with which any detected variant is considered clinically relevant falls within a spectrum, ranging from pathogenic to unrelated to the phenotype.

Germeis et al. introduced a specific customization of ACMG criteria for *SERPING1* to improve variant interpretation of *SERPING1* variants (58). Every detected variant should be assessed with respect to its presence in public, internal and disease-specific databases, population data, computational predictions, *in vivo* and *in vitro* test results, evidence of segregation, and allelic and variant-specific information. The ACMG criteria that are supporting or tolerable must be applied to the abovementioned evidence, whenever possible, resulting in variant classification in one category (i.e., benign, likely benign, VUS, likely pathogenic, and pathogenic) (57). Variants and supporting data should be submitted to public databases upon classification, new evidence that may alter the initial variant assessment and favoring further exchanges between submitters. Online bioinformatic tools can be helpful for determining pathogenicity (e.g., Genetic Variant Interpretation Tool, InterVar, Varsome).

About variants detected in introns, many potential SRE-affecting variants fall into the category of so-called "variants of unknown significance" (104). To distinguish between pathogenic/likely pathogenic mutations and harmless non-splicing-affecting variants, medical geneticists are encouraged to investigate *in vitro* on transcript distribution from patient RNA samples or, although less reliable, to use *in silico* predictions (105). An estimate of exon susceptibility to be skipped or to activate nearby cryptic splice sites can be possible by SRE predictions. An evaluation of their reliability and potential use in clinical diagnostic settings has been developed by Grodecká et al. (71).

## Functional Analysis of Serpin Function

Establishing a dysfunctional C1-INH protein is a prerequisite for further genetic analysis. The common C1-INH function testing should be advantageously completed by an analysis of circulating species for missense variants.

When variants are recognized in an intronic region out of a noncanonical splice sequence, a transcription analysis of an RNA sample extracted from blood nucleated cells is recommended for a functional distinction between a benign and a pathogenic/likely pathogenic variant. Cultured monocytes provide RNA samples of good quality for a downstream analysis; however, the distribution of transcripts could be different from that extracted from blood nucleated cells. A specific approach taking into account a possible NMD of transcripts with premature stop codon should be used to minimize a risk of missing aberrant transcript when evaluating RNA extracted from patients' blood cells (83).

## Structural Analyses With Genotype to Phenotype Correlation; Conserved Positions Among Serpins

C1 Inhibitor controls target proteases as a suicide-substrate where RCL displays the scissile bond Arg<sup>444</sup>-Thr<sup>445</sup> (i.e., P1-P1'; **Figure 3A**). The N-terminal region of RCL is conserved among inhibitory serpins, maintaining proper RCL mobility and loop insertion. However, the sequences adjacent to the cleavage site in the RCL, P6-P4' (i.e., Ala<sup>439</sup>-Val<sup>448</sup>), are highly variable between the different serpins and considered to be a major determinant of serpin specificity (106). In this region, some variants have been recognized as benign/likely benign (e.g., those at the positions Ala<sup>439</sup>, Val<sup>442</sup>, or Val<sup>448</sup>); variants at nonadjacent positions have been characterized as pathogenic because a mutant on the hinge/RCL sequence packs favorably in the loop-inserted latent structure (e.g., those at the positions Ala<sup>436</sup> or Pro<sup>454</sup> located in the proximal and distal hinge, respectively; **Figure 3B**). Importantly because of a lack of the suicide-substrate property of C1-INH, all variants at position Arg<sup>444</sup> are interpreted as pathogenic, inconsistently with commonly used *in silico* predictions.

If the protruding structure of the RCL makes it more accessible for an interaction with target proteases, it also favors the native serpin to be in a stressed thermodynamically metastable conformation M\*. However, polymerized C1-INH represents a stable and low energetic conformation, which can be achieved upon some missense mutations (107). Variants have been recognized as favoring the thermodynamically stable conformation, with susceptibility to spontaneous loop-sheet polymerization or latent serpin species, as described in Section Inheritance of Variants (**Supplementary Table S3**). These variants, whose feature makes it recognized in the class III identified in Section Missense Variants, should be included as HAE-2. **Figure 5** shows an example with the p.(Arg400Cys) variant.

## Frequency of Variants Within Global or Selected Population

Pertaining to the characterization of pathogenic criteria with C1-INH-HAE incidence of 0.00002, submitters are invited, whenever possible, to consider MAF (i.e., MAF in a global/selected population) (57).

**Figure 6** shows a short algorithm displaying the position of biological and genetic analyses within a diagnostic algorithm.

## Contribution to the Field Statement

To our knowledge, this is the largest study taking an overlook of the constellation of *SERPING1* variants found in nearly 1,500 HAE families. This study emphasizes that etiopathogenesis of C1-INH-HAE could be consistently implemented by C1-INH molecular analyses.

Misfolding and polymerization/latentization of the mutated serpins are at the base of a group of conformational diseases collectively known as serpinopathies (25, 29, 107). Likewise, missense mutations in *SERPING1* can cause polymerization/latentization, with an impaired secretion or a failed cleavage by target protease, that lead to C1-INH deficiency and C1-INH-HAE (20, 33, 88, 89).

Recording *SERPING1* genetics together with biological data on serpin function and C1-INH transcripts, for intronic variants out of canonical sequences, should contribute to a high quality value for national registries and for open databases.

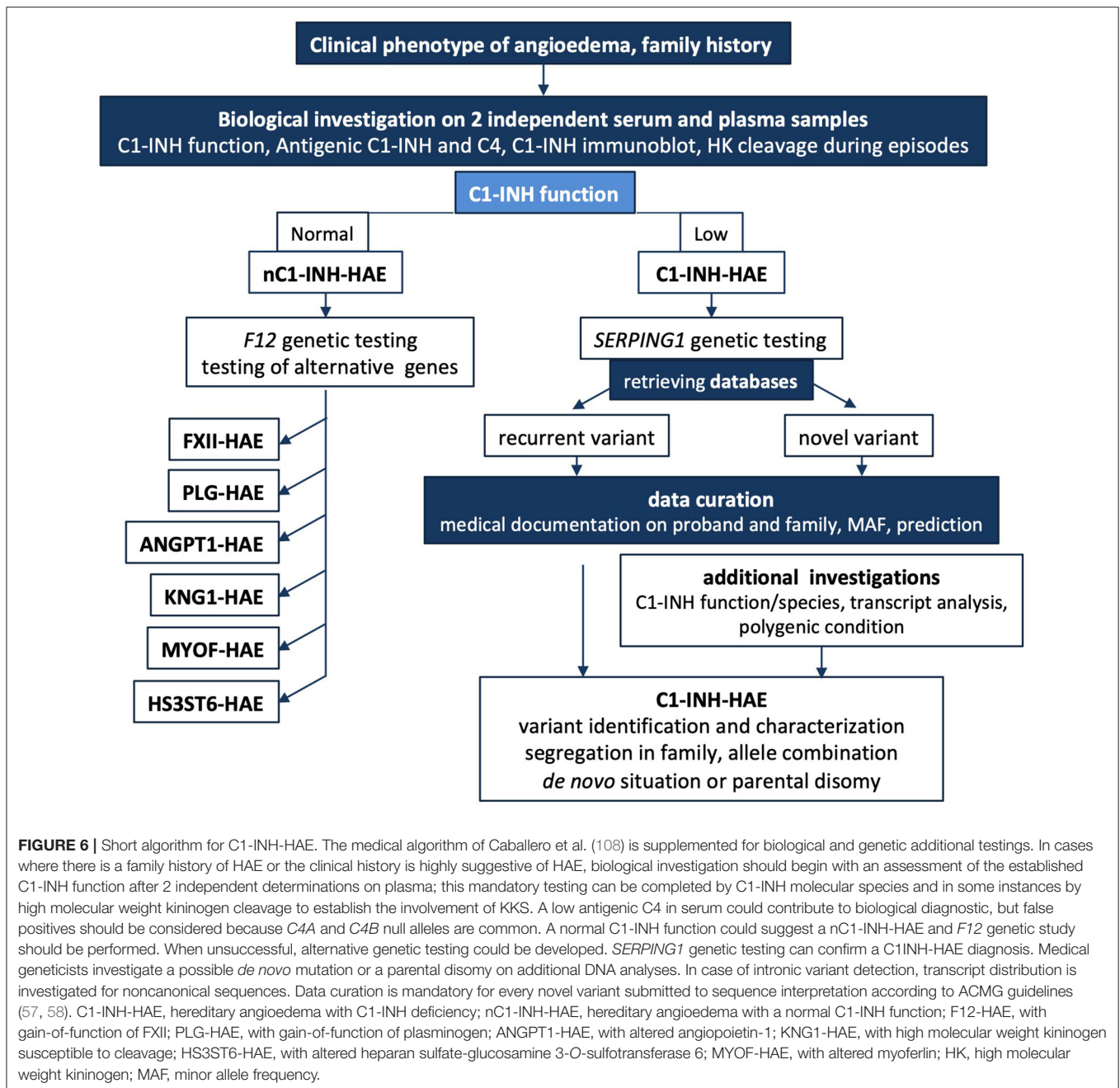
## CONCLUSION AND PERSPECTIVE

Hereditary angioedema due to C1 Inhibitor deficiency has been first characterized as a monogenic disease; however, cumulative arguments on a contribution of additional alleles make clear that clinical variability of C1-INH-HAE is substantially attributed to modifier genes. As suggested by Veronez et al. (36), even if pharmacogenomic associations are very difficult to prove in a rare disease, recommendations will assist the physician for an optimal treatment option for individual patients.

A pathogenic model of C1-INH-HAE has been proposed with a KKS activation in a systemic activation process, where fluid-phase activation of the KKS generates BK, associated with local manifestations after an interaction with locally expressed endothelial kinin receptors (11). The model provides an explanation for why symptoms can occur at multiple sites during an attack and why HAE attacks respond well to modest increases of circulating C1-INH function. The recent observations of multiple allele combinations and of abnormal kinin metabolism are congruent with this model, making C1-INH a strategic component with the participation of additional parameters in patient clinical phenotype.

Functional studies of modulating factors, acting on systemic activation or on activator-bound process, should combine with the discovery of new mutations, with genotype to biological phenotype associations and including a large number of patients.

The document presented here describes the biological and structural features of C1-INH deficiency in relation to groups of *SERPING1* variants. As well as inherited serpinopathies, namely,



**FIGURE 6 |** Short algorithm for C1-INH-HAE. The medical algorithm of Caballero et al. (108) is supplemented for biological and genetic additional testings. In cases where there is a family history of HAE or the clinical history is highly suggestive of HAE, biological investigation should begin with an assessment of the established C1-INH function after 2 independent determinations on plasma; this mandatory testing can be completed by C1-INH molecular species and in some instances by high molecular weight kininogen cleavage to establish the involvement of KKS. A low antigenic C4 in serum could contribute to biological diagnostic, but false positives should be considered because *C4A* and *C4B* null alleles are common. A normal C1-INH function could suggest a nC1-INH-HAE and *F12* genetic study should be performed. When unsuccessful, alternative genetic testing could be developed. *SERPING1* genetic testing can confirm a C1INH-HAE diagnosis. Medical geneticists investigate a possible *de novo* mutation or a parental disomy on additional DNA analyses. In case of intronic variant detection, transcript distribution is investigated for noncanonical sequences. Data curation is mandatory for every novel variant submitted to sequence interpretation according to ACMG guidelines (57, 58). C1-INH-HAE, hereditary angioedema with C1-INH deficiency; nC1-INH-HAE, hereditary angioedema with a normal C1-INH function; F12-HAE, with gain-of-function of FXII; PLG-HAE, with gain-of-function of plasminogen; ANGPT1-HAE, with altered angiotensin-1; KNG1-HAE, with high molecular weight kininogen susceptible to cleavage; HS3ST6-HAE, with altered heparan sulfate-glucosamine 3-O-sulfotransferase 6; MYOF-HAE, with altered myoferlin; HK, high molecular weight kininogen; MAF, minor allele frequency.

$\alpha$ 1-trypsin deficiency, could be considered as ideal candidates for gene therapy, strategies for C1-INH-HAE treatment have been suggested. Based on the argument that HAE should be viewed primarily as a metabolic liver disorder, new therapeutic approaches to C1-INH-HAE have been outlined by Ameratunga et al. (109). Given the very high costs of treating HAE, the authors have considered that gene therapy as curative option may become feasible in the next decade.

In a next future, epigenetics and environmental factors should be considered in the molecular identification of C1-INH-HAE with the characterization of individual severity risk factors as well.

## DATABASES AND BIOINFORMATICS SUPPORTS

- ClinVar: [www.ncbi.nlm.nih.gov/clinvar/](http://www.ncbi.nlm.nih.gov/clinvar/)
- Ensembl: <http://www.ensembl.org>, a centralized resource for geneticists
- Exonic splicing enhancers: ESEfinder release 3.0
- Genetic Variant Interpretation Tool: [http://www.medschool.umaryland.edu/Genetic\\_Variant\\_Interpretation\\_Tool1.html](http://www.medschool.umaryland.edu/Genetic_Variant_Interpretation_Tool1.html)
- Human Splicing Finder, version 3.1: [www.umd.be>hsf](http://www.umd.be>hsf)
- InterVar: <https://wintervar.wglab.org/evds.php>

- LOVD: Leiden Open Variation Database; [databases.lovd.nl/shared/variants/SERPING1](https://databases.lovd.nl/shared/variants/SERPING1)
- Varsome: <https://varsome.com/>, InterVar, wintervar.wglab.org.

## AUTHOR CONTRIBUTIONS

CD, AL-L, AGh, ML-T, SC, DP, MR, TF, CV, JP, and AGE wrote sections or subsections of this manuscript. DP and AGh prepared **Figure 5**. TF and HG prepared **Figure 4**. FP prepared **Supplementary Tables S2, S3**. HG prepared **Supplementary Table S5**. CD, AL-L, AGh, ML-T, SC, DP, MR, TF, CV, JP, and AGE revised the last version of this manuscript. CD supervised the writing of the manuscript and the selection of the figures. All authors contributed to the article and approved the submitted version.

## REFERENCES

- Cicardi M, Banerji A, Bracho F, Malbrán A, Rosenkranz B, Riedl M, et al. Icatibant, a new bradykinin-receptor antagonist, in hereditary angioedema. *N Engl J Med*. (2010) 363:532–41. doi: 10.1056/NEJMx100067
- Nzeako UC, Frigas E, Tremaine WJ. Hereditary angioedema: a broad review for clinicians. *Arch Intern Med*. (2001) 161:2417–29. doi: 10.1001/archinte.161.20.2417
- Schmaier AH. The hereditary angioedema syndromes. *J Clin Invest*. (2019) 129:66–8. doi: 10.1172/JCI125378
- Ghebrehiwet B, Kaplan AP, Joseph K, Peerschke EI. The complement and contact activation systems: partnership in pathogenesis beyond angioedema. *Immunol Rev*. (2016) 274:281–9. doi: 10.1111/imr.12469
- Maas C. Plasmininflammation-An emerging pathway to bradykinin production. *Front Immunol (Lausanne)*. (2019) 10:2046. doi: 10.3389/fimmu.2019.02046
- Irmscher S, Döring N, Halder LD, Jo EAH, Kopka I, Dunker C, et al. Kallikrein cleaves C3 and activates complement. *J Innate Immun*. (2018) 10:94–105. doi: 10.1159/000484257
- Choudhary K, Patel PK, Are VN, Makde RD, Hajela K. Mannose-binding lectin-associated serine protease-1 cleaves plasminogen and plasma fibronectin: prefers plasminogen over known fibrinogen substrate. *Blood Coagul Fibrinolysis*. (2021) 32:504–12. doi: 10.1097/MBC.0000000000001074
- Foley JH. Plasmin(ogen) at the nexus of fibrinolysis, inflammation, and complement. *Semin Thromb Haemost*. (2017) 43:135–42. doi: 10.1055/s-0036-1592302
- Bekassy Z, Lopatko Fagerström I, Bader M, Karpman D. Crosstalk between the renin-angiotensin, complement and kallikrein-kinin systems in inflammation. *Nat Rev Immunol*. (2021) 1–18. doi: 10.1038/s41577-021-00634-8
- Fijen LM, Bork K, Cohn DM. Current and prospective targets of pharmacologic treatment of hereditary angioedema types 1 and 2. *Clin Rev Allergy Immunol*. (2021) 61:66–76. doi: 10.1007/s12016-021-08832-x
- Hofman ZLM, Relan A, Zeerleder S, Drouet C, Zuraw B, Hack CE. Angioedema attacks in patients with hereditary angioedema: local manifestations of a systemic activation process. *J Allergy Clin Immunol*. (2016) 138:359–66. doi: 10.1016/j.jaci.2016.02.041
- Drouet C, Désormeaux A, Robillard J, Ponard D, Bouillet L, Martin L, et al. Metallopeptidase activities in hereditary angioedema: effect of androgen prophylaxis on plasma aminopeptidase P. *J Allergy Clin Immunol*. (2008) 121:429–33. doi: 10.1016/j.jaci.2007.10.048
- Kajdácsi E, Veszeli N, Mezo B, Jandrasics Z, Kohalmi KV, Ferrara AL, et al. Pathways of neutrophil granulocyte activation in hereditary angioedema

## FUNDING

TF and HG were supported by grants NV18-05-00330 (Ministry of Health) and MUNI/A/1099/2019 (Ministry of Education, Youth and Sports, Czech Republic).

## ACKNOWLEDGMENTS

The authors are indebted to Dr. Christine Gaboriaud, Université Grenoble Alpes, CEA, CNRS, IBS Grenoble, for helpful documentation on C1-INH structure.

## SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/falgy.2022.835503/full#supplementary-material>

- with C1 Inhibitor deficiency. *Clin Rev Allergy Immunol*. (2021) 60:383–95. doi: 10.1007/s12016-021-08847-4
- Grymova T, Vlkova M, Soucek P, Hakl R, Nechvatalova J, Slamina P, et al. Neutrophils are dysregulated in patients with hereditary angioedema types I and II in a symptom-free period. *Mediators Inflamm*. (2019) 9515628. doi: 10.1155/2019/9515628
- Csuka D, Varga L, Farkas H, Füst G. Strong correlation of high EBNA-1-IgG levels with edematous attacks involving upper airway mucosa in hereditary angioedema due to C1-inhibitor deficiency. *Mol Immunol*. (2012) 49:649–54. doi: 10.1016/j.molimm.2011.11.005
- López-Lera A, Sánchez Cabo F, Garrido S, Dopazo A, López-Trascasa M. Disease-modifying factors in hereditary angioedema: an RNA expression-based screening. *Orphanet J Rare Dis*. (2013) 8:77. doi: 10.1186/1750-1172-8-77
- Germeis AE, Rijavec M, Veronez CL. Leveraging genetics for hereditary angioedema: a road map to precision medicine. *Clin Rev Allergy Immunol*. (2021) 60:416–28. doi: 10.1007/s12016-021-08836-7
- Rosen FS, Pensky J, Donaldson V, Charache P. Hereditary angioneurotic edema: two genetic variants. *Science*. (1965) 148:957–8. doi: 10.1126/science.148.3672.957
- Cicardi M, Aberer W, Banerji A, Bas M, Bernstein JA, Bork K, et al. Classification, diagnosis, and approach to treatment for angioedema: consensus report from the Hereditary Angioedema International Working Group. *Allergy*. (2014) 69:602–16. doi: 10.1111/all.12380
- Ponard D, Gaboriaud C, Charignon D, Ghannam A, Wagenaar-Bos IGA, Roem D, et al. SERPING1 mutation update: Mutation spectrum and C1 Inhibitor phenotypes. *Hum Mutat*. (2020) 41:38–57. doi: 10.1002/humu.23917
- Henao MP, Kraschewski JL, Kelbel T, Craig TJ. Diagnosis and screening of patients with hereditary angioedema in primary care. *Ther Clin Risk Manag*. (2016) 12:701–11. doi: 10.2147/TCRM.S86293
- Germeis AE, Speletas M. Genetics of hereditary angioedema revisited. *Clin Rev Allergy Immunol*. (2016) 51:170–82. doi: 10.1007/s12016-016-8543-x
- Davis AE. 3rd, Lu F, Mejia P. C1 inhibitor, a multi-functional serine protease. *Inhibitor Thromb Haemost*. (2010) 104:886–93. doi: 10.1160/TH10-01-0073
- Drouet C, Ponard D, Ghannam A. C1 inhibitor. In: Barnum, S, Schein, T, editors. *The Complement FactsBook*. London: Academic Press. p 241–249. (2018). doi: 10.1016/B978-0-12-810420-0.00023-7
- Lomas DA, Belorgey D, Mallya M, Miranda E, Kinghorn KJ, Sharp LK, et al. Molecular mousetraps and the serpinopathies. *Biochem Soc Trans*. (2005) 33:321–30. doi: 10.1042/BST0330321
- Huntington JA, Read RJ, Carrell RW. Structure of a serpin-protease complex shows inhibition by deformation. *Nature*. (2000) 407:923–6. doi: 10.1038/35038119

27. Khan MS, Singh P, Azhar A, Naseem A, Rashid Q, Kabir MA, et al. Serpin inhibition mechanism: a delicate balance between native metastable state and polymerization. *J Amino Acids*. (2011) 2011:606797. doi: 10.4061/2011/606797
28. Wells MJ, Sheffield WP, Blajchman MA. The clearance of thrombin-antithrombin and related serpin-enzyme complexes from the circulation: role of various hepatocyte receptors. *Thromb Haemost*. (1999) 81:325–37doi: 10.1055/s-0037-1614472
29. Goopu B, Lomas DA. Conformational pathology of the serpins: themes, variations, and therapeutic strategies. *Annu Rev Biochem*. (2009) 78:147–76. doi: 10.1146/annurev.biochem.78.082107.133320
30. Windfuhr JP, Alsenz J, Loos M. The critical concentration of C1-esterase inhibitor (C1-INH) in human serum preventing auto-activation of the first component of complement (C1). *Mol Immunol*. (2005) 42:657–63. doi: 10.1016/j.molimm.2004.09.025
31. Wagenaar-Bos IG, Drouet C, Aygören-Pursun E, Bork K, Bucher C, Bygum A, et al. Functional C1-inhibitor diagnostics in hereditary angioedema: assay evaluation and recommendations. *J Immunol Methods*. (2008) 338:14–20. doi: 10.1016/j.jim.2008.06.004
32. Li HH, Busse P, Lumry WR, Frazer-Abel A, Levy H, Steele T, et al. Comparison of chromogenic and ELISA functional C1 inhibitor tests in diagnosing hereditary angioedema. *J Allergy Clin Immunol Pract*. (2015) 3:200–5. doi: 10.1016/j.jaip.2014.08.002
33. Madsen DE, Hansen S, Gram J, Bygum A, Drouet C, Sidemann JJ. Presence of C1-inhibitor polymers in a subset of patients suffering from hereditary angioedema. *PLoS ONE*. (2014) 9:e112051. doi: 10.1371/journal.pone.0112051
34. Hack CE, Relan A, van Amersfoort ES, Cicardi M. Target levels of functional C1-inhibitor in hereditary angioedema. *Allergy*. (2012) 67:123–30. doi: 10.1111/j.1398-9995.2011.02716.x
35. Bouillet L, Ponard D, Rousset H, Cichon S, Drouet C. A case of hereditary angio-oedema type III presenting with C1-inhibitor cleavage and a missense mutation in F12 gene. *Br J Dermatol*. (2007). doi: 10.1111/j.1365-2133.2007.07778.x
36. Veronez CL, Mendes AR, Leite CS, Gomes CP, Grumach AS, Pesquero JB. Hereditary angioedema brazilian study group (GEBRAEH). The panorama of primary angioedema in the Brazilian population. *J Allergy Clin Immunol Pract*. (2021) 9:2293–304.e5.
37. Blanch A, Roche O, Urrutia I, Gamboa P, Fontán G, López-Trascasa M. First case of homozygous C1 inhibitor deficiency. *J Allergy Clin Immunol*. (2006) 118:1330–5. doi: 10.1016/j.jaci.2006.07.035
38. Verpy E, Biasotto M, Brai M, Misiano G, Meo T, Tosi M. Exhaustive mutation scanning by fluorescence-assisted mismatch analysis discloses new genotype-phenotype correlations in angioedema. *Am J Hum Genet*. (1996) 59:308–19.
39. Büyükoztürk S, Eroglu BK, Gelincik A, Uzümcü A, Özşeker F, Colakoglu B, et al. A Turkish family with a novel mutation in the promoter region of the C1 inhibitor gene. *J Allergy Clin Immunol*. (2009) 123:962–4. doi: 10.1016/j.jaci.2008.12.022
40. Kesim B, Uyguner ZO, Gelincik A, Mete Gökmen N, Sin AZ, Karakaya G, et al. The Turkish hereditary angioedema pilot study (TURHAPS): the first Turkish series of hereditary angioedema. *Int Arch Allergy Immunol*. (2011) 156:443–50. doi: 10.1159/000323915
41. Zahedi K, Bissler JJ, Prada AE, Prada JA, Davis 3rd AE. The promoter of the C1 inhibitor gene contains a polypurine-polypyrimidine segment that enhances transcriptional activity. *J Immunol*. (1999) 162:7249–55.
42. Rijavec M, Korošec P, Šilar M, Zidarn M, Miljković J, Košnik M, et al. Hereditary angioedema nationwide study in Slovenia reveals four novel mutations in SERPING1 gene. *PLoS ONE*. (2013) 8:e56712. doi: 10.1371/journal.pone.0056712
43. Kalmár L, Hegedüs T, Farkas H, Nagy M, Tordai A. HAEdb: a novel interactive, locus-specific mutation database for the C1 inhibitor gene. *Hum Mutat*. (2005) 25:1–5. doi: 10.1002/humu.20112
44. Pappalardo E, Caccia S, Suffritti C, Tordai A, Zingale LC, Cicardi M. Mutation screening of C1 inhibitor gene in 108 unrelated families with hereditary angioedema: functional and structural correlates. *Mol Immunol*. (2008) 45:3536–44. doi: 10.1016/j.molimm.2008.05.007
45. Bygum A, Fagerberg CR, Ponard D, Monnier N, Lunardi J, Drouet C. Mutational spectrum and phenotypes in Danish families with hereditary angioedema. *Allergy*. (2011) 66:76–84. doi: 10.1111/j.1398-9995.2010.02456.x
46. López-Lera A, Favier B, Mena de la Cruz R, Drouet C, López-Trascasa M. A new case of homozygous C1-inhibitor deficiency suggests a role for Arg378 in the control of kinin pathway activation. *J Allergy Clin Immunol*. (2010) 126:1307–10.e3. doi: 10.1016/j.jaci.2010.07.037
47. Mete Gökmen N, Gülbahar O, Onay H, Peker Koc Z, Özgül S, Köse T, et al. Deletions in SERPING1 lead to lower C1 Inhibitor function: lower C1 Inhibitor function can predict disease severity. *Int Arch Allergy Immunol*. (2019) 178:50–9. doi: 10.1159/000492583
48. Bafunno V, Divella C, Sessa F, Tiscia GL, Castellano G, Gesualdo L, et al. De novo homozygous mutation of the C1 inhibitor gene in a patient with hereditary angioedema. *J Allergy Clin Immunol*. (2013) 132:748–50.e3. doi: 10.1016/j.jaci.2013.04.006
49. Bors A, Csuka D, Varga L, Farkas H, Tordai A, Füst G, et al. Less severe clinical manifestations in patients with hereditary angioedema with missense C1INH gene mutations. *J Allergy Clin Immunol*. (2013) 131:1708–13. doi: 10.1016/j.jaci.2012.11.015
50. Speletas M, Szilágyi Á, Csuka D, Koutsostathis N, Psarros F, Moldovan D, et al. F12-46C/T polymorphism as modifier of the clinical phenotype of hereditary angioedema. *Allergy*. (2015) 70:1661–4. doi: 10.1111/all.12714
51. Charignon D, Ponard D, de Gennes C, Drouet C, Ghannam A. SERPING1 and F12 combined variants in a hereditary angioedema family. *Ann Allergy Asthma Immunol*. (2018) 121:500–2. doi: 10.1016/j.anaai.2018.05.031
52. Bork K, Zibat A, Ferrari DM, Wollnik B, Schön MP, Wulff K, et al. Hereditary angioedema in a single family with specific mutations in both plasminogen and SERPING1 genes. *J Dtsch Dermatol Ges*. (2020) 18:215–23. doi: 10.1111/ddg.14036
53. Bork K, Wulff K, Steinmüller-Magin L, Braenne I, Staubach-Renz P, Witzke G, et al. Hereditary angioedema with a mutation in the plasminogen gene. *Allergy*. (2018) 73:442–50. doi: 10.1111/all.13270
54. Veronez CL, Aabom A, Martin RP, Filippelli-Silva R, Gonçalves RF, Nicolicht P, et al. Genetic variation of kallikrein-kinin system and related genes in patients with hereditary angioedema. *Front Med (Lausanne)*. (2019) 6:28. doi: 10.3389/fmed.2019.00028
55. Loules G, Parsopoulou F, Zamanakou M, Csuka D, Bova M, González-Quevedo T, et al. Deciphering the genetics of primary angioedema with normal levels of C1 Inhibitor. *J Clin Med*. (2020) 9:3402. doi: 10.3390/jcm9113402
56. Veronez CL, Dias da Silva E, Teixeira PVL, Cagini N, Constantino-Silva RN, Grumach AS, et al. Genetic analysis of hereditary angioedema in a Brazilian family by targeted next generation sequencing. *Biol Chem*. (2016) 397:315–22. doi: 10.1515/hsz-2015-0212
57. Richards S, Aziz N, Bale S, Bick D, Das S, Gastier-Foster J, et al. Standards and guidelines for the interpretation of sequence variants: a joint consensus recommendation of the American College of Medical Genetics and Genomics and the Association for Molecular Pathology. *Genet Med*. (2015) 17:405–24. doi: 10.1038/gim.2015.30
58. Germenis AE, Margaglione M, Pesquero JB, Farkas H, Cichon S, Csuka D, et al. International consensus on the use of genetics in the management of hereditary angioedema. *J Allergy Clin Immunol Pract*. (2020) 8:901–11. doi: 10.1016/j.jaip.2019.10.004
59. Bos IG, Hack CE, Abrahams JP. Structural and functional aspects of C1-inhibitor. *Immunobiology*. (2002) 205:518–33. doi: 10.1078/0171-2985-00151
60. Stoppa-Lyonnet D, Carter PE, Meo T, Tosi M. Clusters of intragenic *Alu* repeats predispose the human C1 inhibitor locus to deleterious rearrangements. *Proc Natl Acad Sci U S A*. (1990) 87:1551–5. doi: 10.1073/pnas.87.4.1551
61. Pappalardo E, Cicardi M, Duponchel C, Carugati A, Choquet S, Agostoni A, et al. Frequent de novo mutations and exon deletions in the C1inhibitor gene of patients with angioedema. *J Allergy Clin Immunol*. (2000) 106:1147–54. doi: 10.1067/mai.2000.110471
62. Duponchel C, Di Rocco C, Cicardi M, Tosi M. Rapid detection by fluorescent multiplex PCR of exon deletions and duplications in the C1 inhibitor

- gene of hereditary angioedema patients. *Hum Mutat.* (2001) 17:61–70. doi: 10.1002/1098-1004(2001)17:1<61::AID-HUMU7>3.0.CO;2-9
63. Roche O, Blanch A, Duponchel C, Fontán G, Tosi M, López-Trascasa M, et al. Hereditary angioedema: the mutation spectrum of SERPING1/C1NH in a large Spanish cohort. *Hum Mutat.* (2005) 26:135–44. doi: 10.1002/humu.20197
  64. Nicolich P, Faria DOS, Martins-Silva L, Maia LSM, Moreno AS, Arruda LK, et al. Gene mapping strategy for Alu elements rearrangements: Detection of new large deletions in the SERPING1 gene causing hereditary angioedema in Brazilian families. *Gene.* (2019) 685:179–85. doi: 10.1016/j.gene.2018.10.084
  65. López-Lera A, Garrido S, Roche O, López-Trascasa M. SERPING1 mutations in 59 families with hereditary angioedema. *Mol Immunol.* (2011) 49:18–27. doi: 10.1016/j.molimm.2011.07.010
  66. Wong WY, Wong H, Au E, Chan E. Identification and mapping of a 2,009-bp DNA deletion in SERPING1 of a hereditary angioedema patient. *Case Rep Genet.* (2019) 7052062. doi: 10.1155/2019/7052062
  67. Loules G, Zamanakou M, Parsopoulou F, Vatsiou S, Psarros F, Csuka D, et al. Targeted next-generation sequencing for the molecular diagnosis of hereditary angioedema due to C1-inhibitor deficiency. *Gene.* (2018) 667:76–82. doi: 10.1016/j.gene.2018.05.029
  68. Grodecká L, Hujová P, Kramárek M, Kršjaková T, Kováčová T, Vondrášková K, et al. Systematic analysis of splicing defects in selected primary immunodeficiencies-related genes. *Clin Immunol.* (2017) 180:3–44. doi: 10.1016/j.clim.2017.03.010
  69. Siddique Z, McPhaden AR, Lappin DE, Whaley K. An RNA splice site mutation in the C1-inhibitor gene causes type I hereditary angio-oedema. *Hum Genet.* (1991) 88:231–2. doi: 10.1007/BF00206079
  70. Andrejević S, Korošec P, Šilar M, Košnik M, Mijanović R, Bonači-Nikolić B, et al. Hereditary angioedema due to C1 Inhibitor deficiency in Serbia: two novel mutations and evidence of genotype-phenotype association. *PLoS ONE.* (2015) 10:e0142174. doi: 10.1371/journal.pone.0142174
  71. Grodecká L, Buratti E, Freiburger T. Mutations of pre-mRNA splicing regulatory elements: are predictions moving forward to clinical diagnostics? *Int J Mol Sci.* (2017) 18:1668. doi: 10.3390/ijms18081668
  72. López-Martínez R, Martínez-Borra J, Fernández-González P, Coto E, Toyos-González P. Hereditary angioedema caused by a novel intronic variant of SERPING1. *Pediatr Allergy Immunol.* (2022) 33:e13681. doi: 10.1111/PAL.13681/v2/response1
  73. Colobran R, Lois S. de la Cruz X, Pujol-Borrell R, Hernández-González M, Guilarte M. Identification and characterization of a novel splice site mutation in the SERPING1 gene in a family with hereditary angioedema. *Clin Immunol.* (2014) 150:143–8. doi: 10.1016/j.clim.2013.11.013
  74. Senapathy P, Shapiro MB, Harris NL. Splice junctions, branch point sites, and exons: sequence statistics, identification, and applications to genome project. *Methods Enzymol.* (1990) 183:252–78. doi: 10.1016/0076-6879(90)83018-5
  75. Roca X, Krainer AR, Eperon IC. Pick one but be quick: 5' splice sites and the problems of too many choices. *Genes Dev.* (2013) 27:129–44. doi: 10.1101/gad.209759.112
  76. Baralle D, Baralle M. Splicing in action: assessing disease causing sequence changes. *J Med Genet.* (2005) 42:737–48. doi: 10.1136/jmg.2004.029538
  77. Obtulowicz K, Ksiazek T, Bogdali A, Dyga W, Czarnobilska E, Juchacz A. Genetic variants of SERPING1 gene in Polish patients with hereditary angioedema due to C1 inhibitor deficiency. *Cent Eur J Immunol.* (2020) 45:301–9. doi: 10.5114/cej.2020.101252
  78. Speletas M, Szilagyi A, Psarros F, Moldovan D, Magerl M, Maria Kompoti M, et al. Hereditary angioedema: Molecular and clinical differences among European populations. *J Allergy Clin Immunol.* (2015) 135:570–3. doi: 10.1016/j.jaci.2014.08.007
  79. Gösswein T, Kocot A, Emmert G, Kreuz W, Martinez-Sauger I, Aygören-Pürsün E, et al. Mutational spectrum of the C1NH (SERPING1) gene in patients with hereditary angioedema. *Cytogenet Genome Res.* (2008) 121:181–8. doi: 10.1159/000138883
  80. Duponchel C, Djenouhat K, Frémeaux-Bacchi V, Monnier N, Drouet C, Tosi M. Functional analysis of splicing mutations and of an exon 2 polymorphic variant of SERPING1/C1NH. *Hum Mutat.* (2006) 27:295–6. doi: 10.1002/humu.9414
  81. Pagani F, Baralle FE. Genomic variants in exons and introns: identifying the splicing spoilers. *Nat Rev Genet.* (2004) 5:389–96. doi: 10.1038/nrg1327
  82. Rodríguez JA, Narváez CF. First Analysis of SERPING1 gene in patients with hereditary angioedema in Colombia reveals two genotypic variants in a highly symptomatic individual. *J Clin Immunol.* (2018) 38:294–9. doi: 10.1007/s10875-018-0491-1
  83. Hujová P, Souček P, Grodecká L, Grombířková H, Ravčuková B, Kuklínek P, et al. Deep intronic mutation in SERPING1 caused hereditary angioedema through pseudoexon activation. *J Clin Immunol.* (2020) 40:435–46. doi: 10.1007/s10875-020-00753-2
  84. Germinen AE, Vatsiou S, Csuka D, Zamanakou M, Farkas H. Deep intronic SERPING1 gene variants: ending one odyssey and starting another? *J Clin Immunol.* (2021) 41:248–50. doi: 10.1007/s10875-020-00887-3
  85. Vatsiou S, Zamanakou M, Loules G, Psarros F, Parsopoulou F, Csuka D, et al. A novel deep intronic SERPING1 variant as a cause of hereditary angioedema due to C1-inhibitor deficiency. *Allergol Int.* (2020) 69:443–9. doi: 10.1016/j.alit.2019.12.009
  86. Kramer J, Rosen FS, Colten HR, Rajczy K, Strunk RC. Transinhibition of C1 inhibitor synthesis in type I hereditary angioneurotic edema. *J Clin Immunol.* (1993) 91:1258–62. doi: 10.1172/JCI116290
  87. Laimer M, Klausegger A, Aberer W, Oende K, Steinhuber M, Lanschuetzer CM, et al. Haploinsufficiency due to deletion within the 3' -UTR of C1-INH-gene associated with hereditary angioedema. *Genet Med.* (2006) 8:249–54. doi: 10.1097/01.gim.0000214302.90076.fa
  88. Haslund D, Ryø LB, Majidi SS, Rose I, Skipper KA, Fryland T, et al. Dominant-negative SERPING1 variants cause intracellular retention of C1 inhibitor in hereditary angioedema. *J Clin Invest.* (2019) 129:388–405. doi: 10.1172/JCI98869
  89. Aulak KS, Eldering E, Hack CE, Lubbers YP, Harrison RA, Mast A, et al. A hinge region mutation in C1-inhibitor (Ala436->Thr) results in nonsubstrate-like behavior and in polymerization of the molecule. *J Biol Chem.* (1993) 268:18088–94. doi: 10.1016/S0021-9258(17)46815-3
  90. Zahedi R, Aulak KS, Eldering E, Davis 3rd AE. Characterization of C1 Inhibitor-Ta A dysfunctional C1INH with deletion of lysine 251. *J Biol Chem.* (1996) 271:24307–12. doi: 10.1074/jbc.271.39.24307
  91. Yasuno S, Ansai O, Hayashi R, Nakamura S, Shimomura Y. Evidence for a dominant-negative effect of a missense mutation in the SERPING1 gene responsible for hereditary angioedema type I. *J Dermatol.* (2021) 48:1243–9. doi: 10.1111/1346-8138.15930
  92. Loli-Ausejo D, López-Lera A, Drouet C, Llluncor M, Phillips-Anglés E, Pedrosa M, et al. In search of an association between genotype and phenotype in hereditary angioedema due to C1-INH deficiency. *Clin Rev Allergy Immunol.* (2021) 61:1–14. doi: 10.1007/s12016-021-08834-9
  93. Caccia S, Suffritti C, Carzaniga T, Berardelli R, Berra S, Martorana V, et al. Intermittent C1-Inhibitor deficiency associated with recessive inheritance: functional and structural insight. *Sci Rep.* (2018) 8:977. doi: 10.1038/s41598-017-16667-w
  94. Bissler JJ, Aulak KS, Donaldson VH, Rosen FS, Cicardi M, Harrison RA, et al. Molecular defects in hereditary angioneurotic edema. *Proc Assoc Am Physicians.* (1997) 109:164–73.
  95. Zahedi R, Bissler JJ, Davis 3rd AE, Andreadis C, Wisnieski JJ. Unique C1 inhibitor dysfunction in a kindred without angioedema II Identification of an Ala443>Val substitution and functional analysis of the recombinant mutant protein. *J Clin Invest.* (1995) 95:1299–305. doi: 10.1172/JCI117780
  96. Zahedi R, MacFarlane RC, Wisnieski JJ, Davis 3rd AE. C1 inhibitor: analysis of the role of amino acid residues within the reactive center loop in target protease recognition. *J Immunol.* (2001) 167:1500–6. doi: 10.4049/jimmunol.167.3.1500
  97. Ghannam A, Sellier P, Defendi F, Favier B, Charignon D, López-Lera A, et al. C1 inhibitor function using contact-phase proteases as target: evaluation of an innovative assay. *Allergy.* (2015) 70:1103–11. doi: 10.1111/all.12657
  98. Guarino S, Perricone C, Guarino MD, Giardina E, Gambardella S, Rosaria D'Apice M, et al. Gonadal mosaicism in hereditary angioedema. *Clin Genet.* (2006) 70:83–85. doi: 10.1111/j.1399-0004.2006.00643.x
  99. Yu T-C, Shyur S-D, Huang L-H, Wen D-C, Li J-S. Paternal mosaicism and hereditary angioedema in a Taiwanese family. *Ann Allergy Asthma Immunol.* (2007) 99:375–9. doi: 10.1016/S1081-1206(10)60557-1
  100. Ebo DG, Van Gasse AL, Sabato V, Bartholomeus E, Reyniers E, Vanbellingen J-F, et al. Hereditary angioedema in 2 sisters due to



- paternal gonadal mosaicism. *J Allergy Clin Immunol Pract.* (2018) 6:277–9. doi: 10.1016/j.jaip.2017.07.002
101. Gianni P, Loules G, Zamanakou M, Kompoti M, Csuka D, Psarros F, et al. Genetic determinants of C1 Inhibitor deficiency angioedema age of onset. *Int Arch Allergy Immunol.* (2017) 174:200–4. doi: 10.1159/000481987
  102. Rijavec M, Košnik M, Andrejević S, Karadža-Lapić L, Grivčeva-Panovska V, Korošec P. The functional promoter F12-46C/T variant predicts the asymptomatic phenotype of C1-INH-HAE. *Clin Exp Allergy.* (2019) 49:1520–2. doi: 10.1111/cea.13470
  103. Vatsiou S, Zamanakou M, Loules G, Gonzáles-Quevedo T, Porenski G, Juchacz A, et al. Aspects of hereditary angioedema genotyping in the era of NGS: The case of F12 gene. *Alergia Astma Immunologia.* (2018) 23:205–10.
  104. Baralle D, Lucassen A, Buratti E. Missed threads. The impact of pre-mRNA splicing defects on clinical practice. *EMBO Rep.* (2009) 10:810–6. doi: 10.1038/embor.2009.170
  105. Buratti E, Baralle M, Baralle FE. From single splicing events to thousands: the ambiguous step forward in splicing research. *Brief Funct Genom.* (2013) 12:3–12. doi: 10.1093/bfpg/els048
  106. Sanrattana W, Maas C, de Maat S. SERPINS-From trap to treatment. *Front Med (Lausanne).* (2019) 6:25. doi: 10.3389/fmed.2019.00025
  107. Roussel BD, Irving JA, Ekeowa UI, Belorgey D, Haq I, Ordóñez A, et al. Unravelling the twists and turns of the serpinopathies. *FEBS J.* (2011) 278:3859–67. doi: 10.1111/j.1742-4658.2011.08201.x
  108. Caballero T, Cabañas R, Pedrosa M. Medical algorithm: management of C1 inhibitor hereditary angioedema. *Allergy.* (2021) 00:1–4. doi: 10.1111/all.15115
  109. Ameratunga R, Bartlett A, McCall J, Steele R, Woon S-T, Katelaris CH. Hereditary angioedema as a metabolic liver disorder: novel therapeutic options and prospects for cure. *Front Immunol.* (2016) 7:547. doi: 10.3389/fimmu.2016.00547

**Conflict of Interest:** AGh was employed by KininX SAS. FP and AGe were employed by CeMIA SA.

The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

**Publisher's Note:** All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2022 Drouet, López-Lera, Ghannam, López-Trascasa, Cichon, Ponard, Parsopoulou, Grombirikova, Freiburger, Rijavec, Veronez, Pesquero and Germenis. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.